

LACORS/HPA Co-ordinated Food Liaison Group Studies: Assessment of the Microbiological Safety of Selected Retail Ready- to-Eat Foods with a focus on *Listeria monocytogenes*.

CL Little[‡], SK Sagoo, IA Gillespie, K Grant, J McLauchlin, and the Food, Water and Environmental Surveillance Network[†].

Department of Gastrointestinal, Emerging and Zoonotic Infections, Health Protection Agency Centre for Infections, 61 Colindale Avenue, London, NW9 5EQ.

‡, Author for correspondence; †, FWES Network comprises Laboratories listed in Annex I

**On behalf of the Local Authorities Co-ordinators of Regulatory Services and the
Health Protection Agency**

Summary

Although listeriosis is a rare cause of human disease in the UK, an increase in the number of cases has been observed since 2001 and notably in persons over 60 years old. This increase prompted this study on the microbiological safety of ready-to-eat foods which included those types potentially linked to cases of listeriosis.

Between May 2006 and April 2007, 6984 ready-to-eat foods were sampled (2168 sliced meats, 1088 sandwiches, 1242 hard cheese, 725 spreadable cheese, 878 butter, 368 probiotic drinks, 515 confectionery products containing fresh or synthetic/imitation cream). Of these, more confectionery products (2.6%), sandwiches (2.0%) and sliced meats (1.1%) were of unsatisfactory quality due to high levels of *Staphylococcus aureus*, *Escherichia coli*, *Listeria monocytogenes* and/or other *Listeria* spp. than other food types (0 - 0.2%).

The food types with the highest prevalence of *L. monocytogenes* were sandwiches (7.0%) and sliced meats (3.7% within shelf-life, 4.2% end of shelf-life). *Listeria monocytogenes* at >100 cfu/g (exceeding EC food safety criteria limit) only occurred in sandwiches (0.4%) and sliced meats (0.7% within shelf-life, 1.0% end of shelf-life).

Contamination with *L. monocytogenes* was more frequent in sandwiches that were pre-packed, had a main sandwich filling of ham, tuna or egg, and/or contained salad ingredients, and in meats that were sliced to order, and stored and/or displayed at >8°C. Acceptable microbiological quality was associated with premises where the management was trained in food hygiene and those that complied with HACCP principles.

This study provides important information about the microbiological safety of ready-to-eat foods and demonstrates that the control of *L. monocytogenes* in such foods, and in particular sandwiches and sliced meats, is essential in order to minimise the potential for this bacterium to be present at levels hazardous to health at the point of consumption. The results of this study will also assist retailers, enforcement officers and policy makers to understand how different food safety practices affect microbiological safety.

Introduction

Human listeriosis is a relatively rare but serious foodborne disease, with high morbidity and mortality in vulnerable populations (e.g. pregnant women and the unborn, newborns, the elderly, and the immunocompromised)¹⁻². It is among the most important causes of death from foodborne infections in industrialised countries³. Since 2000, an increase in the number of listeriosis cases in humans has been observed in several EU countries, including the UK, notably in persons over 60 years old²⁻⁵. Numerous strains of *L. monocytogenes* have contributed to this increase in cases in the UK^{2,6}. A diverse range of ready-to-eat foodstuffs, such as cheeses, butter, meat products and sandwiches, have been implicated in recent reports of transmission of listeriosis^{2,3,7}.

Listeria spp. including *L. monocytogenes* is widely distributed in the environment and is therefore present in a variety of raw food materials. *Listeria monocytogenes* is resistant to diverse environmental conditions, able to grow at refrigeration temperatures and survive high salt concentrations. Its ability to colonise food processing environments is also well recognized⁸. European Commission (EC) Regulation 2073/2005 on microbiological criteria for foodstuffs provides limits for the levels of this bacterium in ready-to-eat food and came into force from January 2006⁹. The limit varies according to the type of consumer, characteristics of the food and information available on the food. Absence of *L. monocytogenes* in 25g is required in some foods, e.g. ready-to-eat foods intended for infants and those for special medical purposes. While for others, such as ready-to-eat foods which do not support growth of *L. monocytogenes* or for which shelf-life assessment has been carried out, the higher limit is 100 cfu/g⁹. Although levels of ≤ 100 cfu/g are considered to be of low risk for human disease in the normal population, the risk is likely to increase for vulnerable populations, e.g. the elderly, and those with poor immune status^{3,10}. The EC Regulation on the hygiene of foodstuffs (Regulation (EC) No. 853/2004)¹¹ provides a risk-based approach to controlling food hygiene. The Regulation requires businesses to implement a written food safety management system based on HACCP principles, and to ensure food handlers are trained or instructed in good hygiene practices.

In 2008, the Scientific Panel on Biological Hazards (BIOHAZ) of the European Food Safety Authority (EFSA) recommended that efforts to reduce risks to human health should focus on risk reduction practices both during the production process of ready-to-eat foods and at home by consumers³. This report recommended the further investigation of listeriosis cases and the generation and analysis of data on ready-to-eat foods where *L. monocytogenes* is most commonly found so as to better assess the risk of foods associated with listeriosis³.

Elderly people are particularly vulnerable to infection. However, the reasons for the increase in listeriosis in this population are not clear. In an attempt to try and understand the increase better, this study focused on the prevalence of *L. monocytogenes* in some ready-to-eat foods. The food types in the study included those that were potentially linked to cases through public health investigations of *L. monocytogenes* in the UK (i.e. sliced meat, sandwiches, cheese, butter)^{2,6}. A range of retail ready-to-eat foods were therefore sampled based upon a 'shopping basket' approach^{12,13} over one year. The shopping basket approach, in which a list of defined food types are collected for sampling, ensures that the range of food products is covered, with a better chance of finding foods of interest at each food premises visited^{12,13}. In order to reflect the potential for deterioration in microbiological quality during shelf-life, pre-packaged sliced meats were tested both immediately after collection and at the end of their shelf life. In addition, information on food labelling instructions relating to storage and use was assessed for all food products.

Materials and Methods

Sample Collection

A total of 6984 food samples collected from retail premises were examined by 31 Official Control Laboratories in the UK between 1 May 2006 and 30 April 2007. This sample set was made up of 2168 sliced meats, 1088 sandwiches, 1242 hard cheese, 725 spreadable cheese, 878 butter, 368 probiotic drinks (i.e. therapeutic milks containing beneficial or 'probiotic' cultures such as lactobacilli and bifidobacteria) and 515 confectionery products containing fresh or synthetic/imitation cream. Registered retail premises lists held by Local Authority (LA) Environmental Health Departments were used to derive an approach to sampling. Retail premises were selected at random from LAs' database of food businesses via a random number generator or every 10th entry and, if suitable, samples were collected. All samples (≥100g) were collected and transported to laboratories by staff from 296 local Environmental Health Departments, involving 51 Local Authority Food Liaison Groups (Annex I), in accordance with the Food Standards Agency Food Law Code of Practice¹⁴ and the Local Authorities Co-ordinators of Regulatory Services (LACORS) guidance on microbiological food sampling¹⁵.

Information on samples was obtained by observation and enquiry and recorded on a standard questionnaire (Annex II). This included relevant information concerning the type of packaging, display/storage temperature and expiry dates. Sampling officers also provided basic information on the premises and its compliance with food hygiene legislation, including: the type of premises, food hygiene inspection category rating, consumers at risk,

confidence in management, level of food hygiene training and compliance with HACCP principles. The food hygiene category rating and information on consumers at risk and confidence in management and control systems (including the application of HACCP-based systems) were based on the risk assessment criteria detailed in the Food Hygiene Inspection Rating Scheme (Annex 5 of the Food Standards Agency Food Law Code of Practice¹⁴).

Sample Examination

Where duplicate pre-packed sliced meat samples were collected, one sample of the two was stored in a monitored laboratory refrigerator at 6±2°C until the end of shelf life (i.e. use-by date), whereupon the samples were examined. All *Listeria* spp. (including *L. monocytogenes*), *Staphylococcus aureus* and *Escherichia coli* were enumerated or their presence sought in accordance with HPA Standard Microbiological Methods for all food samples¹⁶⁻¹⁹. 147 isolates of *L. monocytogenes* were sent to the Food Safety Microbiology Laboratory (FSML), HPA Centre for Infections, for further characterisation. This included sero-typing and amplified fragment length polymorphism (AFLP) as described previously by Doumith *et al.*²⁰ and Guerra *et al.*²¹. Microbiological results were compared to Regulation (EC) 2073/2005 on microbiological criteria for foodstuffs⁹ and Guidelines for the Microbiological Quality of Some Ready-to-eat Foods sampled at the Point of Sale (Table 1)²².

Statistical Analysis

Descriptive and statistical analysis of the data was undertaken using Microsoft Excel and Epi Info version 6.04d. Relative proportions were compared using the Chi squared test (χ^2) and Fisher's exact test. A probability value of less than 5% was defined as significant.

Table 1. Microbiological criteria / guidelines for shopping basket of selected ready-to-eat foods: Key to classification

| Food category | Micro-organism | Microbiological quality (cfu/g unless stated) | | | |
|--|-------------------------------|---|------------------------------------|------------------------------------|---------------------|
| | | Satisfactory | Acceptable | Unsatisfactory | |
| Regulation (EC) No. 2073/2005: Food safety criteria⁹ | | | | | |
| RTE ^a foods placed on the market during their shelf-life | <i>Listeria monocytogenes</i> | ≤10 ² | - | >10 ² | |
| Regulation (EC) No. 2073/2005: Process hygiene criteria (point of production)⁹ | | | | | |
| Butter made from unpasteurised milk | <i>Escherichia coli</i> | <10 | 10 - <10 ² | ≥10 ² | |
| Cheese made from heat treated milk | <i>Escherichia coli</i> | <10 ² | 10 ² - <10 ³ | ≥10 ³ | |
| Fresh cheese made from pasteurised milk | <i>Staphylococcus aureus</i> | <10 | 10 - <10 ² | ≥10 ² | |
| Cheese made from thermised milk and ripened cheese made from pasteurised milk | <i>Staphylococcus aureus</i> | <10 ² | 10 ² - <10 ³ | ≥10 ³ | |
| Cheese made from raw milk | <i>Staphylococcus aureus</i> | <10 ⁴ | 10 ⁴ - <10 ⁵ | ≥10 ⁵ | |
| Guidelines: Microbiological quality at the point of sale²² | | Satisfactory | Acceptable | Unsatisfactory | Unacceptable |
| RTE ^a foods sampled at point of sale | <i>Listeria</i> spp. (Total) | <20 | 20 - <10 ² | ≥10 ² | N/A |
| RTE ^a foods sampled at point of sale | <i>Escherichia coli</i> | <20 | 20 - <10 ² | ≥10 ² | N/A |
| RTE ^a foods sampled at point of sale | <i>Staphylococcus aureus</i> | <20 | 20 - <10 ² | 10 ² - <10 ⁴ | ≥10 ⁴ |

a, RTE, Ready-to-eat; b, N/A, Not applicable.

Results

Listeria monocytogenes contamination of selected foods from retail premises

Overall *L. monocytogenes* was present above 100 cfu/g in 21 of 6984 (0.3%) samples. This level exceeds the food safety criteria in Regulation (EC) No. 2073/2005 for ready-to-eat foods placed on the market during their shelf-life and these samples are thus judged to be legally unsatisfactory⁹. The results were reported to the appropriate food authority, manufacturer and the Food Standards Agency and full investigations were undertaken.

A greater proportion of sandwiches contained *L. monocytogenes* (7.0%) compared to sliced meats within shelf-life (3.7%, $p=0.0003$) or at end of shelf-life (4.2%, $p=0.0174$), confectionery products (0.8%, $p<0.0001$), and hard cheese (0.2%, $p<0.0001$) (Table 2). *Listeria monocytogenes* was not detected in samples of spreadable cheese, butter, or probiotic drinks. However, a higher proportion of sliced meats contained *L. monocytogenes*

at >100 cfu/g (0.7% within shelf-life, 1.0% at end of shelf-life) compared to sandwiches (0.4%), hard cheese (0%), and confectionery products (0%) (Table 2).

Table 2. *Listeria monocytogenes* contamination of selected foods from retail premises

| Product | No. samples | Detection in 25g | | | | Enumeration cfu/g | |
|--|-------------|--------------------|-----------------|--|--|-------------------|-----------------|
| | | Absence (%) | Presence* (%) | | | ≤100 (%) | >100 (%) |
| Sliced meats within shelf-life | 1484 | 1429 (96.3) | 42 (2.8) | | | 3 (0.2) | 10 (0.7) |
| Beef | 218 | 210 (96.3) | 5 (2.3) | | | 1 (0.5) | 2 (0.9) |
| Chicken | 226 | 223 (98.7) | 2 (0.9) | | | 0 | 1 (0.4) |
| Ham | 949 | 909 (95.8) | 31 (3.3) | | | 2 (0.2) | 7 (0.7) |
| Tongue | 91 | 87 (95.6) | 4 (4.4) | | | 0 | 0 |
| Sliced meats at end of shelf-life | 684 | 655 (95.8) | 18 (2.6) | | | 4 (0.6) | 7 (1.0) |
| Beef | 88 | 84 (95.5) | 3 (3.4) | | | 0 | 1 (1.1) |
| Chicken | 121 | 119 (98.3) | 0 | | | 0 | 2 (1.7) |
| Ham | 455 | 432 (94.9) | 15 (3.3) | | | 4 (0.9) | 4 (0.9) |
| Tongue | 20 | 20 (100.0) | 0 | | | 0 | 0 |
| Sandwiches | 1088 | 1012 (93.0) | 63 (5.8) | | | 9 (0.8) | 4 (0.4) |
| Ham | 285 | 265 (93.0) | 17 (5.9) | | | 2 (0.7) | 1 (0.4) |
| Cheese | 204 | 194 (95.1) | 8 (3.9) | | | 2 (1.0) | 0 |
| Tuna | 252 | 227 (90.1) | 20 (7.9) | | | 2 (0.8) | 3 (1.2) |
| Egg mayo | 347 | 326 (94.0) | 18 (5.2) | | | 3 (0.8) | 0 |
| Hard cheese | 1242 | 1240 (99.8) | 2 (0.2) | | | 0 | 0 |
| Cheddar | 802 | 801 (99.8) | 1 (0.2) | | | 0 | 0 |
| Gloucester | 108 | 107 (99.1) | 1 (0.9) | | | 0 | 0 |
| Lancashire | 78 | 78 (100.0) | 0 | | | 0 | 0 |
| Leicester | 254 | 254 (100.0) | 0 | | | 0 | 0 |
| Confectionery products with | 515 | 511 (99.2) | 4 (0.8) | | | 0 | 0 |
| Fresh cream | 392 | 388 (99.0) | 4 (1.0) | | | 0 | 0 |
| Synthetic cream | 123 | 123 (100.0) | 0 | | | 0 | 0 |
| Butter | 878 | 878 (100.0) | 0 | | | 0 | 0 |
| Spreadable cheese | 725 | 725 (100.0) | 0 | | | 0 | 0 |
| Probiotic drinks | 368 | 368 (100.0) | 0 | | | 0 | 0 |

*Detected in 25g & <10 cfu/g

92 (62.6%) of the 147 *L. monocytogenes* isolates were serogroup 1/2a, 37 (21.1%) 4b, 21 (14.3%) 1/2b, and 3 (2.0%) 1/2c. Overall, 16 different *L. monocytogenes* subtypes were obtained with the 1/2a IX, 1/2a VII, 1/2b II, 4b I, 4b IV subtypes recovered from 74.8% of these (Table 3): 16 different subtypes were obtained from sandwiches, 9 from sliced meats, 2 from hard cheese, and 2 from confectionary products.

Table 3. Subtypes of *L. monocytogenes* isolated from sliced meats, sandwiches, hard cheese and confectionery products

| Typing character (Serotype/AFLP*) | No. Samples (%) | Sliced meats (No. samples) | Sandwiches (No. samples) | Hard cheese (No. samples) | Confectionery products (No. samples) |
|-----------------------------------|-----------------|--|--|---------------------------|---|
| 1/2a I | 1 (0.7) | | 1 (egg mayo) | | |
| 1/2a II | 2 (1.4) | | 2 (ham salad) | | |
| 1/2a III | 7 (4.7) | 5 (ham) | 2 (egg mayo (1), ham salad (1)) | | |
| 1/2a VI | 2 (1.4) | | 2 (tuna salad) | | |
| 1/2a VII | 19 (12.9) | 10 (beef (1), ham (9)) | 9 (cheese salad (1), egg mayo (4), ham (1), ham salad (2), tuna salad (1)) | | |
| 1/2a IX | 53 (36.1) | 25 (beef (2), chicken (1), ham (21), tongue (1)) | 27 (cheese salad (3), egg mayo (8), ham salad (3), tuna salad (13)) | 1 (cheddar) | |
| 1/2a XI | 2 (1.4) | | 2 (egg mayo) | | |
| 1/2a XIV | 5 (3.4) | 3 (beef (1), ham (2)) | 2 (ham (1), ham salad (1)) | | |
| 1/2a XVIII | 1 (0.7) | | 1 (ham) | | |
| 1/2b II | 16 (10.8) | 9 (beef (2), chicken (3), ham (4)) | 4 (egg mayo (1), ham salad (1), tuna salad (2)) | | 3 (dairy cream selection, chocolate éclair, vanilla slices) |
| 1/2b IV | 5 (3.4) | 3 (ham) | 1 (cheese salad) | 1 (gloucester) | |
| 1/2c VII | 3 (2.0) | 2 (ham) | 1 (egg mayo) | | |
| 4b I | 11 (7.5) | 6 (ham (4), tongue (2)) | 4 (cheese salad (2), egg mayo (1), tuna salad (1)) | | 1 (dairy cream selection) |
| 4b IV | 11 (7.5) | 8 (beef (2), ham (6)) | 3 (cheese salad (1), ham (1), ham salad (1)) | | |
| 4b V | 7 (4.7) | | 7 (cheese salad (1), ham salad (3), tuna (1), tuna salad (2)) | | |
| 4b IX | 2 (1.4) | | 2 (ham) | | |
| Total | 147 | 71 | 70 | 2 | 4 |

*, Amplified fragment length polymorphism

Microbiological quality of sliced meats

Of the 1484 sliced meats sampled, 64% were ham, 15.2% chicken, 14.7% beef, and 6.1% tongue (Table 4). Overall 1.1% of sliced meats were of unsatisfactory microbiological quality due to high levels of *E. coli* (range of 1.0×10^2 – 1.6×10^3 cfu/g), *S. aureus* (range of 1.6 – 2.0×10^2 cfu/g), *L. monocytogenes* (range of 1.2×10^2 – 8.0×10^5 cfu/g), and other *Listeria* spp. (*L. innocua* and *L. welshimeri*, range of 3.3×10^2 – 1.8×10^3 cfu/g). More sliced beef samples were of unsatisfactory quality (2.3%) than other meat types (0.4 – 1.1%) (Table 4). However, both *Listeria* spp. and *L. monocytogenes* were detected in fewer sliced chicken

samples (2.7% and 1.3%, respectively) compared to other meat types (6.6–9.2% *Listeria* spp. ($p=0.0058$); 3.7–4.4% *L. monocytogenes* ($p=0.0357$)) (Table 4).

Forty six percent (684/1484) of samples collected were duplicate pre-packed sliced meat samples, of which one sample of the two were examined at the end of shelf life (i.e. use-by date) (Table 4). There was no significant difference in the proportion of sliced meats of unsatisfactory microbiological quality at the end of shelf-life (1.4%) compared to those tested within shelf-life (1.1%) ($p>0.05$) (Table 4). Likewise, a similar proportion of samples at the end of shelf-life contained *Listeria* spp. and *L. monocytogenes* (7.0% and 4.3%, respectively) compared to those within shelf-life (6.6% and 3.7%, respectively) (Table 4).

Eighty percent of sliced meats sampled were not cooked on site (Table 4). There was no significant difference in the microbiological quality of sliced meats, nor to the presence of *Listeria* spp. and *L. monocytogenes*, in relation to whether the meat was cooked on or off the premises (Table 4) ($p>0.05$).

Eighty two percent of the sliced meats collected were pre-packed whilst the remainder (18%) were sliced to order (Table 4). More samples that were sliced to order contained *Listeria* spp. and *L. monocytogenes* (10.8% and 3.8%, respectively) compared to those that were pre-packed (5.8% ($p=0.0043$) and 3.0%, respectively) (Table 4). There was no significant difference in the proportion of sliced meats of unsatisfactory microbiological quality and whether samples were pre-packed or sliced to order (1.1-1.2%, $p>0.05$).

Of the pre-packed meat samples, 67.4% were modified atmosphere packed (MAP), 6.5% were vacuum packed (VP), 8.3% were packed in a normal atmosphere, and for 7.8% this information was not recorded (Table 4). Significantly more VP samples (3.0%) were of unsatisfactory microbiological quality compared with MAP (0.8%) or normal atmosphere packed samples (1.0%) ($p=0.0254$). However, a higher proportion of samples that were packed in a normal atmosphere contained *Listeria* spp. and *L. monocytogenes* (11.8% and 8.9%, respectively) compared to those that were either MAP packed (4.8% and 3.3%, respectively, $p=0.0086$) or VP packed (7.0% and 4.0%, respectively, $p>0.05$) (Table 4).

Ninety six percent of all sliced meat samples sold pre-packed were not packaged on the premises of sale (Table 4). Although the total numbers were low, more pre-packed samples packaged on the premises were of unsatisfactory microbiological quality (2.3%) than those samples packaged elsewhere (1.1%). The place of packaging was not significantly associated with contamination rates for *Listeria* spp. and *L. monocytogenes* (packaged on-site, 4.6% and 2.3%, respectively; packaged off-site, 5.8% and 3.9%, respectively) ($p>0.05$, Table 4).

Sixty seven percent of sliced meats had a pack size of 100 - <200g (Table 4). More samples of pack size ≥ 300 g (2.6%) were of unsatisfactory microbiological quality compared with those of smaller pack sizes (0.8-1.5%). A significantly higher proportion of samples of

Table 4. Microbiological quality of different sliced meats

| Sliced meat details | Total No. Samples n=1484 (%) | | No. Samples Unsatisfactory n = 17 (%) | | Samples with all <i>Listeria</i> spp. <i>Listeria</i> spp. n = 99 (%) | | Samples with <i>L. monocytogenes</i> <i>L. monocytogenes</i> n = 55 (%) | |
|--|---------------------------------|--------|--|-------|--|--------|--|--------|
| Type | | | | | | | | |
| Beef | 218 | (14.7) | 5 | (2.3) | 20 | (9.2) | 8 | (3.7) |
| Chicken | 226 | (15.2) | 1 | (0.4) | 6 | (2.7) | 3 | (1.3) |
| Ham | 949 | (63.9) | 10 | (1.1) | 67 | (7.1) | 40 | (4.2) |
| Tongue | 91 | (6.1) | 1 | (1.1) | 6 | (6.6) | 4 | (4.4) |
| Pre-packed & tested at end of shelf-life (n=684) | | | | | | | | |
| Beef | 88 | (12.9) | 1 | (1.1) | 7 | (8.0) | 4 | (4.6) |
| Chicken | 121 | (17.7) | 2 | (1.6) | 3 | (2.5) | 2 | (1.6) |
| Ham | 455 | (66.5) | 7 | (1.5) | 38 | (8.4) | 23 | (5.1) |
| Tongue | 20 | (2.9) | 0 | | 0 | | 0 | |
| Cooked on site | | | | | | | | |
| Yes | 304 | (20.5) | 3 | (1.0) | 23 | (7.6) | 10 | (3.3) |
| No | 1180 | (79.5) | 14 | (1.2) | 76 | (6.4) | 45 | (3.8) |
| Packaging | | | | | | | | |
| Pre-packed | 1216 | (81.9) | 14 | (1.2) | 70 | (5.8) | 47 | (3.8) |
| Sliced to order | 268 | (18.1) | 3 | (1.1) | 29 | (10.8) | 8 | (3.0) |
| Pre-packed product (n=1216) | | | | | | | | |
| Vacuum packed* | 200 | (16.5) | 6 | (3.0) | 14 | (7.0) | 8 | (4.0) |
| Modified atmosphere packed† | 820 | (67.4) | 7 | (0.8) | 39 | (4.8) | 27 | (3.3) |
| Normal atmosphere packed | 101 | (8.3) | 1 | (1.0) | 12 | (11.8) | 9 | (8.9) |
| Not recorded | 95 | (7.8) | 0 | | 5 | (5.3) | 3 | (3.2) |
| Pre-packed product packed on site (n=1216) | | | | | | | | |
| Yes | 44 | (3.6) | 1 | (2.3) | 2 | (4.6) | 1 | (2.3) |
| No | 1172 | (96.4) | 13 | (1.1) | 68 | (5.8) | 46 | (3.9) |
| Pack size: | | | | | | | | |
| <100g | 206 | (13.9) | 3 | (1.5) | 8 | (3.9) | 5 | (2.4) |
| 100 - <200g | 991 | (66.8) | 10 | (1.0) | 64 | (6.5) | 34 | (3.4) |
| 200 - <300g | 126 | (8.5) | 1 | (0.8) | 10 | (7.9) | 6 | (4.8) |
| ≥300g | 78 | (5.3) | 2 | (2.6) | 11 | (14.1) | 9 | (11.5) |
| Not recorded | 83 | (5.5) | 1 | (1.2) | 6 | (7.2) | 1 | (1.2) |
| Temperature stored/displayed | | | | | | | | |
| ≤ 5°C | 684 | (46.1) | 12 | (1.7) | 63 | (9.2) | 32 | (4.6) |
| > 5 - ≤ 8°C | 680 | (45.8) | 4 | (0.6) | 25 | (3.6) | 17 | (2.5) |
| > 8°C (range: 9–20°C) | 65 | (4.4) | 1 | (1.5) | 8 | (12.3) | 5 | (7.7) |
| Not recorded | 55 | (3.7) | 0 | | 3 | (5.5) | 1 | (1.8) |
| Country of origin: | | | | | | | | |
| Belgium | 30 | (2.0) | 0 | | 1 | (3.3) | 0 | |
| Brazil | 7 | (0.5) | 0 | | 0 | | 0 | |
| Denmark | 10 | (0.7) | 0 | | 0 | | 0 | |
| Germany | 14 | (0.9) | 0 | | 0 | | 0 | |
| Italy | 3 | (0.2) | 0 | | 0 | | 0 | |
| Poland | 2 | (0.2) | 0 | | 0 | | 0 | |
| Republic of Ireland | 23 | (1.6) | 0 | | 3 | (13.0) | 1 | (4.4) |
| South America | 15 | (1.0) | 0 | | 0 | | 0 | |
| UK | 1088 | (73.3) | 13 | (1.2) | 72 | (6.6) | 43 | (4.0) |
| Produce of EC | 5 | (0.3) | 0 | | 0 | | 0 | |
| Not known | 287 | (19.3) | 4 | (1.4) | 23 | (8.0) | 11 | (3.8) |

* Vacuum packaging is essentially the evacuation of air from a package that is then hermetically sealed.

† Modified atmosphere packaging is the removal of air and replacement by a strictly controlled gaseous mixture comprising of carbon dioxide, oxygen and/or nitrogen.

pack size ≥300g contained *Listeria* spp. and *L. monocytogenes* (14.1% and 11.5%, respectively) compared to those of smaller pack sizes (3.9-7.9% *Listeria* spp. (p=0.0154);

2.4-4.8% *L. monocytogenes* ($p=0.0022$) (Table 4). Furthermore, the proportion of samples with *Listeria* spp. decreased as the pack size decreased ($\geq 300\text{g}$, 14.1%, 200 - $<300\text{g}$, 7.9%; 100 - $<200\text{g}$, 6.5%; $<100\text{g}$, 3.9% (chi-square trend $p=0.0026$) (Table 4).

In general storage temperatures for cooked meats should be as low as possible, i.e. $\leq 5^{\circ}\text{C}$. Higher temperatures (up to $\leq 8^{\circ}\text{C}$) may be used with an appropriate shelf-life^{23,24}. At the time of sampling, the air temperature between the sliced meats on display or storage was $\leq 5^{\circ}\text{C}$ for 46.1% of samples, and between $>5 - \leq 8^{\circ}\text{C}$ for a further 45.8% of samples (Table 4). The display or storage temperature of the sliced meats on display had no significant effect on the microbiological quality of the meat with regard to samples of unsatisfactory microbiological quality ($p>0.05$) (Table 4). A higher proportion of samples that were displayed or stored at $\leq 5^{\circ}\text{C}$ contained *Listeria* spp. and *L. monocytogenes* (9.2% and 4.6%, respectively) compared to those displayed/stored at $>5^{\circ}\text{C}$ to $\leq 8^{\circ}\text{C}$ (3.6% and 2.5%, respectively) ($p<0.0001$) but not those at $>8^{\circ}\text{C}$ (12.3% and 7.7%, respectively) ($p>0.05$) (Table 4). *Listeria monocytogenes* at $>10^2$ cfu/g was present in 1.0% (7/684) of meat samples displayed/stored at $\leq 5^{\circ}\text{C}$.

It is illegal to sell food older than its use-by date. This date was recorded on the packaging for 94.2% (1146 of 1216) of the pre-packed sliced meat samples. Based on the use-by date, 17.4% (200 of 1146) of samples collected had remaining shelf lives ranging from 0 to 5 days, 35.2% (404) had 6 to 10 days, 46.7% (537) had 11 days or more, and 0.4% (5) had expired (4 by 1 day, 1 by 3 days) and therefore were not in compliance with the UK food labelling regulations of 1996. *L. monocytogenes* at >100 cfu/g was more likely to be found in sliced meats with a remaining shelf life of between 0 to 5 days (1.5%, 3/200) or 6 to 10 days (1.0%, 4/404) than in meats with more than 10 days left (0.4%, 2/537).

Seventy three percent of sliced meat samples were identified as being of United Kingdom origin; 7.4% of samples were identified as originating from another EU or non-EU country. The place of origin was not known for 19.3% of samples.

Microbiological quality of sandwiches

Of the 1088 sandwiches, 31.8% of samples contained egg mayonnaise as the main sandwich filling, 26.2% ham, 23.2% tuna, and 18.8% cheese (Table 5). Overall 2.0% of sandwiches were of unsatisfactory microbiological quality due to high levels of *E. coli* (range of $1.2 \times 10^2 - 6.9 \times 10^3$ cfu/g), *S. aureus* (range of $1.0 \times 10^2 - 1.8 \times 10^3$ cfu/g), *L. monocytogenes* (range of $2.2 \times 10^2 - 1.2 \times 10^3$ cfu/g), and other *Listeria* spp. (*L. innocua*, range of $3.7 \times 10^2 - 2.6 \times 10^4$ cfu/g; *L. welshimeri*, range of $1.8 \times 10^2 - 1.7 \times 10^3$ cfu/g). More ham sandwiches were of unsatisfactory quality (3.5%) compared to other filling types (1.0 – 2.0%) (Table 5). Both *Listeria* spp. and *L. monocytogenes* was detected less

frequently in cheese sandwiches (9.8% and 4.9%, respectively) compared to other filling types (12.7–14.7% *Listeria* spp.; 6.1–9.9% *L. monocytogenes*) (Table 5).

Table 5. Microbiological quality of different sandwiches

| Sandwich details | Total No. Samples n=1088 (%) | No. Samples Unsatisfactory n = 22 (%) | Samples with all <i>Listeria</i> spp. n = 139 (%) | Samples with <i>L. monocytogenes</i> n = 76 (%) |
|--|---------------------------------|--|--|--|
| Sandwich filling | | | | |
| Ham | 285 (26.2) | 10 (3.5) | 42 (14.7) | 20 (7.0) |
| Cheese | 204 (18.8) | 2 (1.0) | 20 (9.8) | 10 (4.9) |
| Tuna | 252 (23.2) | 3 (1.2) | 33 (13.1) | 25 (9.9) |
| Egg mayonnaise | 347 (31.8) | 7 (2.0) | 44 (12.7) | 21 (6.1) |
| Sandwich filling with salad ingredients | | | | |
| Yes | 760 (69.9) | 17 (2.2) | 114 (15.0) | 66 (8.9) |
| No | 328 (30.1) | 5 (1.5) | 25 (7.6) | 10 (3.1) |
| Packaging | | | | |
| Pre-packed | 888 (81.6) | 17 (1.9) | 128 (14.4) | 72 (8.1) |
| Made to order | 200 (18.4) | 5 (2.5) | 11 (5.5) | 4 (2.0) |
| Pre-packed sandwiches, stored/displayed (n=888) | | | | |
| ≤ 5°C | 520 (58.6) | 7 (1.3) | 80 (15.3) | 48 (9.2) |
| >5 - ≤ 8°C | 279 (31.4) | 8 (2.8) | 37 (13.2) | 19 (6.8) |
| > 8°C (range: 9–24°C) | 58 (6.5) | 1 (1.7) | 4 (8.6) | 3 (5.1) |
| Not recorded | 31 (3.5) | 1 (3.2) | 7 (22.6) | 2 (6.4) |

Seventy percent of sandwiches collected also contained salad ingredients (Table 5). The proportion of samples of unsatisfactory microbiological quality that contained salad was similar (2.2%) to those that did not (1.5%). However, significantly more sandwiches with salad ingredients contained *Listeria* spp. (15.0%) and *L. monocytogenes* (8.9%) compared with those without salad ingredients (7.6%, *Listeria* spp. ($p=0.0007$); 3.1% *L. monocytogenes* ($p=0.0006$)) (Table 5).

Eighty two percent of sandwiches collected were pre-packed whilst 18.4% were made to order (Table 5). There was no significant difference in the proportion of sandwiches of unsatisfactory microbiological quality and the packaging used (1.9-2.5%, $p>0.05$). However, a significantly higher proportion of samples that were pre-packed contained *Listeria* spp. and *L. monocytogenes* (14.4% and 8.1%, respectively) compared to those that were made to order (5.5% and 2.0%, respectively) ($p=0.0004$) (Table 5).

The British Sandwich Association recommends that sandwiches should be delivered and stored or retailed at 5°C and never higher than 8°C²⁵. At the time of sampling, 58.6% of pre-packed sandwiches were displayed or stored at ≤5°C, and a further 31.4% at between >5 - ≤8°C (Table 5). The display or storage temperature of the pre-packed sandwiches on display or storage had no significant effect on the microbiological quality of the sandwiches with regard to samples of unsatisfactory microbiological quality ($p>0.05$) nor to the proportion of sandwiches that contained *Listeria* spp. and *L. monocytogenes* ($p>0.05$) (Table 5).

Thirty six percent (391/1088) of sandwiches that contained salad ingredients were pre-packed and stored or displayed at $\leq 5^{\circ}\text{C}$. Of these:

- 17.3% and 10.7% contained *Listeria* spp. and *L. monocytogenes* respectively, and
- 1.2% and 0.8% contained *Listeria* spp. and *L. monocytogenes* respectively at $>10^2$ cfu/g.

The use by date was recorded on the packaging for 83.0% (826 of 888) of the prepacked sandwich samples. Based on the use-by date, 92.4% (764 of 826) of samples collected had remaining shelf lives ranging from 0 to 2 days, 7.3% (60) had 3 to 4 days, and 0.3% (2) had expired (1 by 1 day and 1 by 2 days) and therefore were not in compliance with the UK food labelling regulations of 1996. *L. monocytogenes* was more likely to be found in sandwiches with a remaining shelf life of 2 days or less (7.5%, 57/764) than in sandwiches with more than 2 day lefts (16.7, 10/60) ($p=0.0229$). The four samples that contained *L. monocytogenes* at >100 cfu/g all had 2 days of shelf life remaining.

Microbiological quality of hard cheese

Most of the 1242 hard cheeses sampled were either cheddar (64.6%) or leicester (20.4%) varieties (Table 6). Overall 0.2% of hard cheese samples were of unsatisfactory microbiological quality due to high levels of *E. coli* (range of 2.4×10^3 – 1.1×10^4 cfu/g). A small proportion of cheddar (0.3%) and leicester (1.3%) cheese samples were of unsatisfactory microbiological quality. Significantly, a higher proportion of lancashire cheese samples contained *Listeria* spp. (7.7%) compared with other hard cheese varieties (1.2-2.8%) ($p=0.0143$). *L. monocytogenes* was recovered from a 0.1% of cheddar and 0.9% of gloucester cheeses samples (Table 6).

Sixty one percent of hard cheese samples collected were made using pasteurised milk (Table 6). More cheese samples made from unpasteurised milk were of unsatisfactory microbiological quality (5.9%) and contained *Listeria* spp. (5.9%) compared to pasteurised milk cheeses (0.3% unsatisfactory quality; 2.4% *Listeria* spp.) (Table 6).

Ninety one percent of the hard cheeses collected were pre-packed and 55.7% had a pack size of between 200 and 300g (Table 6). Significantly, more samples that were cut to order (1.7%) were of unsatisfactory quality compared to those that were pre-packed (0.1%) ($p=0.0248$) (Table 5). *Listeria* spp. and *L. monocytogenes* were only detected in pre-packed cheeses (Table 6).

Eighty five percent of hard cheese samples were identified as being of United Kingdom origin; 1.4% of samples were identified as originating from another EU country. This place of origin was not known for 13.6% of samples.

Table 6. Microbiological quality of different hard cheeses

| Hard cheese details | Total No. Samples n = 1242 (%) | No. Samples Unsatisfactory n = 3 (%) | Samples with all <i>Listeria</i> spp. n = 34 (%) | Samples with <i>L. monocytogenes</i> n = 2 (%) |
|--------------------------|-----------------------------------|---|---|---|
| Type | | | | |
| Cheddar | 802 (64.6) | 2 (0.3) | 22 (2.7) | 1 (0.1) |
| Gloucester | 108 (8.7) | 0 | 3 (2.8) | 1 (0.9) |
| Lancashire | 78 (6.3) | 1 (1.3) | 6 (7.7) | 0 |
| Leicester | 254 (20.4) | 0 | 3 (1.2) | 0 |
| Made from | | | | |
| Pasteurised milk | 758 (61.0) | 2 (0.3) | 18 (2.4) | 2 (0.3) |
| Unpasteurised milk | 17* (1.4) | 1† (5.9) | 1‡ (5.9) | 0 |
| Not known | 467 (37.6) | 0 | 15 (3.2) | 0 |
| Packaging | | | | |
| Pre-packed | 1125 (90.6) | 1 (0.1) | 34 (3.0) | 2 (0.2) |
| Cut to order | 117 (9.4) | 2 (1.7) | 0 | 0 |
| Pack size | | | | |
| <100g | 28 (2.3) | 0 | 1 (3.6) | 0 |
| 100 - <200g | 296 (23.8) | 3 (1.0) | 4 (1.4) | 0 |
| 200 - <300g | 692 (55.7) | 0 | 24 (3.5) | 2 (0.3) |
| ≥300g | 160 (12.9) | 0 | 3 (1.9) | 0 |
| Not recorded | 66 (5.3) | 0 | 2 (3.0) | 0 |
| Stored/kept | | | | |
| ≤ 8°C | 1100 (88.6) | 2 (0.2) | 27 (2.5) | 1 (0.1) |
| > 8°C (range: 9–29°C) | 92 (7.4) | 1 (1.1) | 3 (3.3) | 1 (1.1) |
| Not recorded | 50 (4.0) | 0 | 4 (8.0) | 0 |
| Country of origin | | | | |
| Belgium | 2 (0.2) | 0 | 0 | 0 |
| Canada | 1 (0.1) | 0 | 0 | 0 |
| Germany | 6 (0.5) | 0 | 0 | 0 |
| Netherlands | 2 (0.2) | 0 | 0 | 0 |
| Republic of Ireland | 4 (0.3) | 0 | 0 | 0 |
| UK | 1057 (85.1) | 2 (0.2) | 30 (2.8) | 2 (0.2) |
| Not known | 170 (13.6) | 1 (0.6) | 4 (2.4) | 0 |

*Cheese types made from unpasteurised milk included: cheddar (15), lancashire (1), leicester (1)

† lancashire cheese, ‡ cheddar cheese

Eighty nine percent of cheese samples were stored or displayed at or below 8°C (Table 6). More cheese samples (1.1%) that were stored above 8°C were of unsatisfactory microbiological quality compared to those stored below 8°C (0.2%). Of cheese samples that were stored above 8°C, 3.3% contained *Listeria* spp. and 1.1% *L. monocytogenes* which was similar to those stored below 8°C (2.5% *Listeria* spp.; 0.1% *L. monocytogenes*).

Microbiological quality of butter

Eighty percent of all the 878 butter samples collected were salted, 10.2% were lightly salted and 7.4% were unsalted: for 2.4% of samples, this information was not known (Table 7). Overall, 0.2% of butter samples were of unsatisfactory microbiological quality due to high levels of *E. coli* (range of 3.5×10^2 – 3.4×10^4 cfu/g). Only samples of unsalted butter were of unsatisfactory microbiological quality (3.1%, $p=0.0057$) or contained *Listeria* spp. (detected in 25g) (4.6%, $p=0.0004$) but not *L. monocytogenes* (Table 7).

Ninety seven percent of butter samples were made from pasteurised milk (Table 7). None of the seven samples made from unpasteurised milk were of unsatisfactory microbiological quality or contained *Listeria* spp. Only 0.2% of cheeses made from pasteurised milk were of unsatisfactory microbiological quality and 0.4% contained *Listeria* spp. (Table 7).

All samples were pre-packed and 90.0% were a pack size of between 200 and <300g (Table 7). Butter of unsatisfactory microbiological quality was only found in pack sizes 100 - <200g (4.6%) and 200 - <300g (0.1%). *Listeria* spp. was only found in pack sizes of 200 to 300g (Table 7). Most samples were stored or displayed at $\leq 8^{\circ}\text{C}$ (89.9%) (Table 7). Butter samples of unsatisfactory quality (0.2%) were only found in butter stored or displayed at $\leq 8^{\circ}\text{C}$. *Listeria* spp. was also only found in butter stored or displayed at $\leq 8^{\circ}\text{C}$ (0.4%) (Table 7).

Most butter samples were marked as from the UK (62.7%) or Denmark (17.5%) (Table 6), and 0.4% of these were of unsatisfactory quality. *Listeria* spp. was only recovered in samples of butter originating from the UK (0.6%) (Table 7).

Table 7. Microbiological quality of butter

| Butter details | Total No. Samples n = 878 (%) | | No. Samples Unsatisfactory n = 2 (%) | | Samples with all <i>Listeria</i> spp.* n = 3 (%) | |
|---------------------------------------|----------------------------------|--------|--|-------|--|-------|
| Type | | | | | | |
| Salted (1-2%) | 703 | (80.0) | 0 | | 0 | |
| Lightly salted (<1%) | 89 | (10.2) | 0 | | 0 | |
| Unsalted (0%) | 65 | (7.4) | 2 | (3.1) | 3 | (4.6) |
| Not known | 21 | (2.4) | 0 | | 0 | |
| Made from | | | | | | |
| Pasteurised milk | 853 | (97.1) | 2 | (0.2) | 3 | (0.4) |
| Unpasteurised milk | 7 | (0.8) | 0 | | 0 | |
| Not known | 18 | (2.1) | 0 | | 0 | |
| Pack size | | | | | | |
| <100g | 3 | (0.3) | 0 | | 0 | |
| 100 - <200g | 22 | (2.5) | 1 | (4.6) | 0 | |
| 200 - <300g | 798 | (90.9) | 1 | (0.1) | 3 | (0.4) |
| $\geq 300\text{g}$ | 17 | (2.0) | 0 | | 0 | |
| Not recorded | 38 | (4.3) | 0 | | 0 | |
| Stored/kept | | | | | | |
| $\leq 8^{\circ}\text{C}$ | 789 | (89.9) | 2 | (0.2) | 3 | (0.4) |
| $> 8^{\circ}\text{C}$ (range: 9–20°C) | 52 | (5.9) | 0 | | 0 | |
| Not recorded | 37 | (4.2) | 0 | | 0 | |
| Country of origin | | | | | | |
| Channel Islands | 3 | (0.3) | 0 | | 0 | |
| Denmark | 154 | (17.5) | 0 | | 0 | |
| France | 17 | (1.9) | 0 | | 0 | |
| Germany | 16 | (1.8) | 0 | | 0 | |
| Lithuania | 1 | (0.1) | 0 | | 0 | |
| New Zealand | 63 | (7.2) | 0 | | 0 | |
| Poland | 7 | (0.8) | 0 | | 0 | |
| Republic of Ireland | 29 | (3.3) | 0 | | 0 | |
| UK | 550 | (62.7) | 2 | (0.4) | 3 | (0.6) |
| Not known | 38 | (4.3) | 0 | | 0 | |

Microbiological quality of spreadable cheese

Of the 725 pre-packed spreadable cheeses sampled, 74.8% were classified as cheese spread and 25.2% as cream cheese. Most (91.3%) were unflavoured (Table 8). Ninety one percent of the spreadable cheeses collected were of pack size 100-<200g (52.0%) or 200-<300g (36.3%), and 89.7% were stored or displayed at $\leq 8^{\circ}\text{C}$ (Table 8).

Of the cheese spread samples, 0.2% were of unsatisfactory microbiological quality due to high levels of *E. coli* at 4.6×10^2 cfu/g. *Listeria* spp. was recovered from 0.4% of the cheese spreads and 0.6% of the cream cheese samples (Table 8).

The spreadable cheeses samples were produced in nine countries. The majority of samples were produced in Germany (24.1%), Belgium (21.1%), the UK (18.8%), or Denmark (13.3%) (Table 8). Of the samples of UK origin, 0.7% were of unsatisfactory microbiological quality. *Listeria* spp. was recovered from 2.3% and 1.1% of spreadable cheeses originating from the Republic of Ireland and Germany, respectively (Table 8).

Table 8. Microbiological quality of different spreadable cheeses

| Spreadable cheese details | Total No. Samples n = 725 (%) | No. Unsatisfactory n = 1 (%) | Samples n = 3 (%) | Samples with all <i>Listeria</i> spp. n = 3 (%) |
|---|----------------------------------|---------------------------------|----------------------|---|
| Type | | | | |
| Cheese spread | 542 (74.8) | 1 (0.2) | 2 (0.4) | |
| Cream cheese | 183 (25.2) | 0 | 1 (0.6) | |
| Flavour | | | | |
| Unflavoured | 662 (91.3) | 1 (0.2) | 3 (0.5) | |
| Flavoured (chives, garlic, herbs, pepper) | 63 (8.7) | 0 | 0 | |
| Pack size | | | | |
| <100g | 37 (5.1) | 0 | 0 | |
| 100 - <200g | 377 (52.0) | 0 | 1 (0.3) | |
| 200 - <300g | 263 (36.3) | 0 | 1 (0.4) | |
| $\geq 300\text{g}$ | 14 (1.9) | 0 | 0 | |
| Not recorded | 34 (4.7) | 1 (2.9) | 1 (2.9) | |
| Stored/kept: | | | | |
| $\leq 8^{\circ}\text{C}$ | 650 (89.7) | 1 (0.2) | 2 (0.3) | |
| $> 8^{\circ}\text{C}$ (range: 9–20°C) | 33 (4.6) | 0 | 0 | |
| Not recorded | 42 (5.7) | 0 | 1 (2.4) | |
| Country of origin | | | | |
| Belgium | 153 (21.1) | 0 | 0 | |
| Denmark | 97 (13.3) | 0 | 0 | |
| France | 49 (6.8) | 0 | 0 | |
| Germany | 175 (24.1) | 0 | 2 (1.1) | |
| Italy | 2 (0.3) | 0 | 0 | |
| Poland | 11 (1.5) | 0 | 0 | |
| Republic of Ireland | 43 (6.0) | 0 | 1 (2.3) | |
| Spain | 2 (0.3) | 0 | 0 | |
| UK | 136 (18.8) | 1 (0.7) | 0 | |
| Not known | 57 (7.8) | 0 | 0 | |

Microbiological quality of probiotic drinks

Of the 368 probiotic drinks sampled, 47.8% were unflavoured and 52.2% were flavoured. The majority of samples were of pack size 100-<200g (35.1%) or ≥300g (36.1%). Ninety percent (90.2%) were stored or displayed at ≤8°C (Table 9).

No samples were of unsatisfactory microbiological quality. *Listeria* spp. was recovered from 1.7% of the unflavoured and 0.5% of the flavoured drinks (Table 9).

Probiotic drinks collected were produced in nine countries. The majority were produced in the UK (22.3%), France (14.7%) or Belgium (12.0%) (Table 9). *Listeria* spp. was only recovered from 1.1% of probiotic drink samples. *Listeria* spp. was recovered in 5.0% of samples that originated from Poland, 4.6% from Belgium and 1.9% from France (Table 9).

Table 9. Microbiological quality of pre-packed probiotic drinks

| Probiotic drinks details | Total No. Samples n = 368 (%) | | Samples with all <i>Listeria</i> spp. n = 4 (%) | |
|---|----------------------------------|--------|--|-------|
| Type | | | | |
| Unflavoured | 176 | (47.8) | 3 | (1.7) |
| Flavoured (i.e. strawberry, raspberry, orange, mixed fruit) | 192 | (52.2) | 1 | (0.5) |
| Pack size | | | | |
| <100g | 46 | (12.5) | 0 | |
| 100 - <200g | 129 | (35.1) | 1 | (0.8) |
| 200 - <300g | 6 | (1.6) | 0 | |
| ≥300g | 133 | (36.1) | 2 | (1.5) |
| Not recorded | 54 | (14.7) | 1 | (1.8) |
| Stored/kept | | | | |
| ≤ 8°C | 332 | (90.2) | 4 | (1.2) |
| > 8°C (range: 9–19°C) | 8 | (2.2) | 0 | |
| Not recorded | 28 | (7.6) | 0 | |
| Country of origin | | | | |
| Austria | 20 | (5.4) | 0 | |
| Belgium | 44 | (12.0) | 2 | (4.6) |
| France | 54 | (14.7) | 1 | (1.9) |
| Germany | 16 | (4.4) | 0 | |
| Netherlands | 27 | (7.3) | 0 | |
| Poland | 20 | (5.4) | 1 | (5.0) |
| Republic of Ireland | 4 | (1.1) | 0 | |
| Spain | 6 | (1.6) | 0 | |
| UK | 82 | (22.3) | 0 | |
| Not known | 95 | (25.8) | 0 | |

Microbiological quality of confectionery products

Of all the 515 confectionery products sampled, 76.1% contained fresh cream and 23.9% synthetic or imitation cream (Table 10). Amongst those containing fresh cream, 2.6% were of unsatisfactory microbiological quality due to high levels of *E. coli* (range of 2.2×10^2 – 7.0×10^3 cfu/g), *S. aureus* (1.0 – 3.0×10^2 cfu/g), or *Listeria* spp. (*L. innocua* 1.7×10^3 cfu/g); this finding was not statistically significant ($p>0.05$). More samples of confectionery products

with fresh cream contained *Listeria* spp. (3.3%) than those with synthetic cream (0.8%). *L. monocytogenes* were only recovered from products with fresh cream (1.0%) (Table 10).

Of confectionery products collected, 61.7% were pre-packed whilst 38.3% were open (Table 10). Significantly more samples that were open were of unsatisfactory microbiological quality (4.6%) compared with those that were pre-packed (0.3%) ($p=0.0010$). A higher proportion of samples that were open contained *Listeria* spp. and *L. monocytogenes* (5.1% and 1.0%, respectively) compared to those that were pre-packed (1.3% and 0.6%, respectively). This finding was statistically significant for *Listeria* spp. ($p=0.0124$) but not for *L. monocytogenes* (Table 10).

Of all the confectionery products, 72.4% were stored or displayed at $\leq 8^{\circ}\text{C}$ (Table 10), and amongst these, 2.4% were of unsatisfactory microbiological quality. More samples that were displayed or stored at $\leq 8^{\circ}\text{C}$ contained *Listeria* spp. and *L. monocytogenes* (3.2% and 1.1%, respectively) compared to those displayed or stored at $> 8^{\circ}\text{C}$ (1.0% and 0%, respectively) (Table 10).

Ninety seven percent of the confectionery products were produced in the UK (Table 10), and of these 1.8% were of unsatisfactory microbiological quality. *Listeria* spp. and *L. monocytogenes* was only recovered from confectionery products of UK origin (2.8% and 0.8%, respectively) (Table 10). However, it should be noted that the proportion of samples produced outside the UK and examined was small and that no statistical conclusions should be drawn from these results.

Table 10. Microbiological quality of confectionery products

| Confectionery product details | Total Samples n = 515 (%) | No. Samples Unsatisfactory n = 10 (%) | Samples with all <i>Listeria</i> spp. n = 14 (%) | Samples with <i>L. monocytogenes</i> n = 4 (%) |
|--|------------------------------|---|---|---|
| Containing | | | | |
| Fresh cream | 392 (76.1) | 10 (2.6) | 13 (3.3) | 4 (1.0) |
| Synthetic/imitation cream | 123 (23.9) | 0 | 1 (0.8) | 0 |
| Packaging | | | | |
| Pre-packed | 318 (61.7) | 1 (0.3) | 4 (1.3) | 2 (0.6) |
| Open | 197 (38.3) | 9 (4.6) | 10 (5.1) | 2 (1.0) |
| Stored/kept: | | | | |
| $\leq 8^{\circ}\text{C}$ | 373 (72.4) | 9 (2.4) | 12 (3.2) | 4 (1.1) |
| $> 8^{\circ}\text{C}$ (range: 10–26°C) | 104 (20.2) | 0 | 1 (1.0) | 0 |
| Not recorded | 38 (7.4) | 1 (2.6) | 1 (2.6) | 0 |
| Country of origin | | | | |
| Denmark | 2 (0.4) | 0 | 0 | 0 |
| Germany | 2 (0.4) | 0 | 0 | 0 |
| Republic of Ireland | 1 (0.2) | 0 | 0 | 0 |
| UK | 499 (96.9) | 9 (1.8) | 14 (2.8) | 4 (0.8) |
| Not known | 11 (2.1) | 1 (9.1) | 0 | 0 |

Microbiological quality of selected foods from retail premises in relation to product labelling, storage information, shelf-life and durability dates

Amongst all the 5558 pre-packed foods sampled, 43.4% had no instructions on the packaging/label relating to shelf-life after opening (e.g. consume within 3 days), whilst 39.8% did. This information was not recorded for 16.8% of samples. Most foods sampled (87.0%) had storage temperature information present on the packaging or label. Of these, 64.8% advised to keep the product refrigerated with 22.2% specifying a refrigeration temperature (Table 11). Where a particular storage temperature was provided on the packaging for 1234 samples, 79.0% specified a temperature between 0 and $\leq 5^{\circ}\text{C}$ and 30.0% between >5 and $\leq 8^{\circ}\text{C}$ (Table 11). Samples that had no storage temperature information (1.7%) or instructions to store at a particular temperature (1.2%) were more likely to be of unsatisfactory microbiological quality compared to those that instructed to 'keep refrigerated' (0.3%) ($p < 0.0001$) (Table 11). However, more samples that gave specific a storage temperature of $\leq 5^{\circ}\text{C}$ contained *Listeria* spp. and *L. monocytogenes* (8.0% and 5.0%, respectively) compared to those that specified to 'keep refrigerated' or provided no information (*Listeria* spp. 3.0-6.1% ($p < 0.0001$, $p = 0.0465$), *L. monocytogenes* 1.1-3.8% ($p < 0.0001$, $p = 0.0460$)) (Table 11).

Table 11. Microbiological quality of pre-packed foods from retail premises in relation to product labelling, storage information, shelf-life and durability dates

| | Total Samples n = 5558 (%) | No. No. Samples Unsatisfactory n = 36 (%) | Samples with all <i>Listeria</i> spp. n = 246 (%) | Samples with <i>L. monocytogenes</i> n = 123 (%) |
|--|----------------------------------|--|---|--|
| Storage temperature information on packaging/label | | | | |
| Keep refrigerated | 3603 (64.8) | 11 (0.3) | 108 (3.0) | 39 (1.1) |
| Particular temperature given | 1234 (22.2) | 15 (1.2) | 96 (7.8) | 60 (4.8) |
| 0 - ≤5°C | 975 (79.0) | 15 (1.5) | 78 (8.0) | 49 (5.0) |
| >5 - ≤8°C | 259 (30.0) | 0 | 18 (6.9) | 11 (4.2) |
| No temperature indicated | 519 (9.3) | 9 (1.7) | 32 (6.1) | 20 (3.8) |
| Not recorded | 202 (3.7) | 1 (0.5) | 10 (4.9) | 4 (2.0) |
| Storage, shelf-life, durability date details | | | | |
| Easily visible & clearly legible | | | | |
| Yes | 5120 (92.1) | 29 (0.6) | 228 (4.4) | 115 (2.2) |
| No | 278 (5.0) | 5 (1.7) | 8 (2.9) | 4 (1.4) |
| Not recorded | 160 (2.9) | 2 (1.3) | 10 (6.3) | 4 (2.5) |
| In black type on white background | | | | |
| Yes | 3450 (62.1) | 19 (0.6) | 162 (4.7) | 74 (2.1) |
| No | 1708 (30.7) | 11 (0.6) | 61 (3.6) | 38 (2.2) |
| Not recorded | 400 (7.2) | 6 (1.5) | 23 (5.6) | 11 (2.3) |
| Font size of type/print | | | | |
| 10 point | 2468 (44.4) | 12 (0.5) | 101 (4.1) | 55 (2.2) |
| 8 point | 1033 (18.6) | 7 (0.7) | 46 (4.5) | 26 (2.5) |
| Other (6,12,14,16,18 point) | 1456 (26.2) | 7 (0.5) | 71 (4.9) | 27 (1.8) |
| Not recorded | 601 (10.8) | 10 (1.7) | 28 (4.6) | 15 (2.5) |

The storage, shelf-life information and durability dates were easily visible and clearly legible as judged by the sampling officer for 92.1% of all the samples collected (Table 11). A significantly lower proportion of samples where these details were easily visible or legible were of unsatisfactory microbiological quality (0.6%) as compared to those where it was not (1.7%; $p=0.0285$) (Table 11). For 62.1% of the samples, the storage, shelf-life information and durability dates were printed or written in black type on a white background, with 44.4% having a 10 point font size (Table 11). In 2.6% of the packages this information was printed in yellow and white, and in 2.9% using 6 point font size, which is a format that is difficult to read (as shown here).

There was no significant difference in the proportion of samples of unsatisfactory microbiological quality and the colour of print type and background used (0.6%) or the font size used (0.5-0.7%) ($p>0.05$). There was also no significant difference in the proportion of samples that had *Listeria* spp. and *L. monocytogenes* present and storage, shelf-life and durability marking details (Table 11) ($p>0.05$).

Microbiological quality of selected foods in relation to premises details

Amongst all of the samples, 64% were collected from supermarkets, and 21.1% from convenience shops. The remaining 15.0% of samples were collected from bakeries (3.5%), delicatessens (3.2%), butchers (2.2%), farm shops/markets (1.9%), sandwich bars (1.7%), market stalls (0.2%) and other premises (2.3%) (Table 12).

Food businesses are categorised from A to E in accordance with risk assessment criteria of the Food Hygiene Inspection Rating Scheme (Annex 5 of the FSA Food Law Code of Practice¹⁴). Premises in Category A receive the highest minimum inspection frequency. Category ratings are based findings at inspection, and take into account the type of food and method of handling, compliance with food hygiene legislation, and confidence in management and control systems. Over half (54.0%) of samples were collected from premises categorised as Category C in the Food Hygiene Inspection Rating Scheme (Table 12). Significantly, more samples of unsatisfactory microbiological quality were collected from Category A premises (13.0%) than premises in other categories (0.4-1.1%) ($p=0.0011$). A significantly higher proportion of samples containing *Listeria* spp. and *L. monocytogenes* were from Category A premises (17.3% and 8.7%, respectively) than premises in other categories (4.2-6.6% *Listeria* spp. ($p<0.0001$); 2.0-2.6% *L.monocytogenes* ($p=0.0183$)) (Table 12).

As part of the Food Hygiene Inspection Rating Scheme, businesses receive a 'Consumer at Risk' score based on the number of consumers likely to be at risk if there is a failure of food hygiene and procedures¹⁴. This score ranges from 0 to 15, with a score of 0 indicating very few consumers at risk. Eighty seven percent of samples were obtained from

Table 12. Microbiological quality of selected foods collected from retail premises

| Premises details | Total Samples n= 6299 (%) | No. Satisfactory n = 55 (%) | No. Unsatisfactory n = 55 (%) | Samples with all <i>Listeria</i> spp. n = 296 (%) | Samples with <i>L. monocytogenes</i> n = 137 (%) |
|--|----------------------------------|--------------------------------|----------------------------------|--|---|
| Premises Type | | | | | |
| Bakery | 217 (3.5) | 12 (5.5) | 18 (8.3) | 8 (3.7) | |
| Butcher | 140 (2.2) | 3 (2.1) | 17 (12.1) | 6 (4.2) | |
| Convenience shop | 1331 (21.1) | 15 (1.1) | 92 (6.9) | 55 (4.1) | |
| Delicatessen | 201 (3.2) | 4 (1.9) | 5 (2.5) | 1 (0.5) | |
| Farm shop/market | 119 (1.9) | 4 (3.3) | 7 (5.8) | 2 (1.7) | |
| Market shop/stall | 15 (0.2) | 0 | 0 | 0 | |
| Sandwich bar | 106 (1.7) | 3 (2.8) | 12 (11.3) | 6 (5.7) | |
| Supermarket delicatessen | 203 (3.2) | 0 | 6 (3.0) | 2 (1.0) | |
| Supermarket pre-packed | 3820 (60.7) | 13 (0.3) | 134 (3.5) | 55 (1.4) | |
| Other (Service station, cash & carry, greengrocer, village shop) | 147 (2.3) | 1 (0.7) | 5 (3.4) | 2 (1.4) | |
| Inspection Rating Category | | | | | |
| Category | Minimum Frequency of Inspection | | | | |
| A | At least every 6 months | 23 (0.4) | 3 (13.0) | 4 (17.3) | 2 (8.7) |
| B | At least every 12 months | 499 (7.9) | 3 (0.6) | 33 (6.6) | 13 (2.6) |
| C | At least every 18 months | 3403 (54.0) | 39 (1.1) | 152 (4.5) | 69 (2.0) |
| D | At least every 2 years | 1141 (18.1) | 4 (0.4) | 61 (5.4) | 29 (2.5) |
| E | Alternative enforcement strategy | 707 (11.2) | 4 (0.6) | 30 (4.2) | 17 (2.4) |
| Not recorded | | 526 (8.4) | 2 (0.4) | 16 (3.0) | 7 (1.3) |
| Consumers at Risk Score | | | | | |
| 0 (Very few) | 121 (1.9) | 0 | 7 (5.6) | 5 (4.1) | |
| 5 (Few) | 4368 (69.3) | 49 (1.1) | 227 (5.2) | 110 (2.5) | |
| 10 (Intermediate) | 1099 (17.5) | 2 (0.2) | 35 (3.2) | 12 (1.1) | |
| 15 (Substantial) | 41 (0.7) | 0 | 1 (2.4) | 1 (2.4) | |
| Not recorded | 670 (10.6) | 4 (0.6) | 26 (3.9) | 9 (1.3) | |
| Confidence in Management Score | | | | | |
| 0 (High) | 443 (7.0) | 1 (0.2) | 15 (3.3) | 8 (1.8) | |
| 5 (Moderate) | 2173 (34.5) | 12 (0.6) | 78 (3.6) | 23 (1.1) | |
| 10 (Some) | 2398 (38.1) | 30 (1.3) | 132 (5.5) | 68 (2.8) | |
| 20 (Little) | 549 (8.7) | 8 (1.5) | 42 (7.6) | 27 (4.9) | |
| 30 (None) | 30 (0.5) | 0 | 3 (10.0) | 2 (6.7) | |
| Not recorded | 706 (11.2) | 4 (0.6) | 26 (3.7) | 9 (1.3) | |
| Compliant with principles of HACCP[†] (EC No. 852/2004, Article 5) | | | | | |
| Yes | 4434 (70.4) | 35 (0.8) | 189 (4.3) | 40 (0.9) | |
| No | 926 (14.7) | 13 (1.4) | 61 (6.6) | 74 (7.9) | |
| Not recorded | 939 (14.9) | 7 (0.8) | 46 (4.9) | 23 (2.4) | |
| Manager Food Hygiene Training | | | | | |
| Received training & attended | 4735 (75.1) | 40 (0.8) | 204 (4.3) | 89 (1.9) | |
| - Foundation | 3031 (64.0) | 34 (1.1) | 147 (4.9) | 72 (2.4) | |
| - Intermediate | 882 (18.6) | 4 (0.5) | 29 (3.3) | 11 (1.3) | |
| - Advanced | 195 (4.1) | 1 (0.5) | 10 (5.1) | 1 (0.5) | |
| - Other course (Company training, MLC HACCP, C&G) | 333 (7.0) | 1 (0.3) | 12 (3.6) | 3 (0.9) | |
| - Not recorded | 294 (6.3) | 0 | 6 (2.0) | 2 (0.7) | |
| No training | 697 (11.1) | 8 (1.1) | 51 (7.3) | 33 (4.7) | |
| Not recorded | 867 (13.8) | 7 (0.8) | 41 (4.7) | 15 (1.7) | |

†, Hazard Analysis and Critical Control Points

premises with a Consumer at Risk score of 5 (69.3%) or 10 (17.5%) (Table 12). A significantly greater proportion of samples collected from premises with a Consumer at Risk score of 5 (1.1%) were of unsatisfactory microbiological quality as compared to those from

premises with higher Consumer at Risk scores (0.2%, intermediate to substantial numbers) ($p=0.0014$) (Table 12). A higher proportion of samples contained *Listeria* spp. and *L. monocytogenes* were recovered from premises with Consumer at Risk score of 0 to 5 (5.2% and 2.7%, respectively) compared to those with scores of 10-15 (3.4% and 1.2%, respectively) ($p<0.0024$) (Table 12). Furthermore, the proportion of samples with *Listeria* spp. decreased as the Consumer at Risk score increased (0 (very few consumers at risk), 5.6%; 5 (few), 5.2%; 10 (intermediate), 3.2%; 15 (substantial), 2.4%) (chi-square trend $p=0.0046$) (Table 12).

In accordance with the Food Hygiene Inspection Rating Scheme, businesses are allocated a score of 0 to 30 based on Confidence in Management/Control Systems, with 0 indicating a high level of high confidence¹⁴. Seventy three percent of samples were collected from premises that had a Confidence in Management¹⁴ score 5 (34.5%, moderate confidence in management/control systems) and 10 (38.1%, some confidence in management/control systems) (Table 12). The proportion of unsatisfactory samples was significantly higher from premises where there was no, little or some confidence in management (1.3%) compared to premises where there was moderate or high confidence in management (0.5%) ($p=0.0027$) (Table 12). A significantly higher proportion of samples from premises where there was no, little or some confidence in management contained *Listeria* spp. and *L. monocytogenes* (5.8% and 3.3%, respectively) compared to premises where there was moderate or high confidence in management (3.6% and 1.1%, respectively) ($p<0.0001$) (Table 12). Furthermore, the proportion of samples with *Listeria* spp. decreased as confidence in management increased (0 (high confidence), 3.3%; 5 (moderate), 3.6%; 10 (some), 5.5%; 20 (little), 7.6%, 30 (none), 10.0%) (Chi-square trend $p<0.0001$) (Table 12).

Seventy percent of samples collected were from premises that, in view of the sampling officer, complied with HACCP requirements as provided in Article 5 of Regulation (EC) No. 852/2004¹⁰ (Table 12). Samples collected from premises that did not comply with this requirement were more likely to be of unsatisfactory microbiological quality (1.4%) than those collected from premises that did (0.8%) (Table 12). A significantly higher proportion of samples collected from premises that did not comply with the HACCP requirements contained *Listeria* spp. and *L. monocytogenes* (6.6% and 4.3%, respectively) compared to those that did (4.3% and 1.7%, respectively) ($p=0.0035$) (Table 12).

Seventy five percent of samples were collected from premises whose managers had received food hygiene training (Table 12). Samples collected from premises with managers that had not received food hygiene training were more likely to be of unsatisfactory microbiological quality (1.1%) than those with managers who had received training in food hygiene (0.8%) (Table 12). Significantly, a higher proportion of samples collected from premises with managers that had not received food hygiene training contained *Listeria* spp.

and *L. monocytogenes* (7.3% and 4.7%, respectively) than those with managers with training in food hygiene (4.3% and 1.9%, respectively) ($p=0.0010$) (Table 12).

Discussion

This study has shown that overall the vast majority (99.1%) of selected ready-to-eat foods on retail sale in the UK were of satisfactory or acceptable microbiological quality according to criteria in Regulation (EC) No. 2073/2005 (as amended)⁹ and published microbiological guidelines²². Of the foods examined, a higher proportion of confectionery products (2.6%), sandwiches (2.0%) and sliced meats (1.1%) were of unsatisfactory quality due to high levels of *S. aureus*, *E. coli*, *L. monocytogenes* and/or other *Listeria* spp. than hard cheese (0.2%), butter (0.2%), spreadable cheese (0.2%) and probiotic drinks (0%). High *E. coli*, *S. aureus* and *Listeria* levels may indicate problems with production hygiene, and/or that the temperature of these foods in storage or on display was inadequate to prevent bacterial growth.

The prevalence of *L. monocytogenes* observed in sandwiches (7.0%) and sliced meats (3.7% within shelf-life, 4.2% end of shelf-life) was higher than that found in all other ready-to-eat foods examined (0 – 0.8%). Furthermore, *L. monocytogenes* at >100 cfu/g (food safety criteria limit)⁸ only occurred in sandwiches (0.4%) and sliced meats (0.7% within shelf-life, 1.0% end of shelf-life). Full investigations of these incidents were undertaken by the appropriate food authorities, manufacturers and the UK Food Standards Agency. In Wales during 2005 to 2006, retail sandwiches were found to have a similar prevalence of *L. monocytogenes* (5.2%) and exceeded 100 cfu/g in 0.3% of sandwiches²⁶. Previously in 1996 Wilson²⁷ reported *L. monocytogenes* present at >100 cfu/g in 0.7% of retail sandwiches in Northern Ireland whereas in Ireland in 2002, 11% of retail sandwiches contained *L. monocytogenes*, and 0.3% at >100 cfu/g²⁸. Sandwiches supplied to hospitals and residential or care homes in the UK during 2005-2006 were also found to have a similar rate of contamination of *L. monocytogenes* (7.5%) to that reported in the present study²⁹. While *L. monocytogenes* is the species of concern, the presence of any *Listeria* spp. in food is an indication of poor hygiene conditions and possible *L. monocytogenes* contamination. Significant risk factors identified with the presence of *Listeria* spp. and *L. monocytogenes* in sandwiches sampled in the present study were if: pre-packed; contained ham, tuna or egg mayonnaise as the main sandwich filling; and/or contained salad ingredients. A range of food types have been associated with transmission of listeriosis, these food types have also included sandwiches^{2,3}. The microbiological quality of ingredients incorporated in to sandwiches is, therefore, of importance as is for all ready-to-eat products. The British Sandwich Association recommends a target level of <10 cfu/g of *L. monocytogenes* in sandwiches at production, and that the presence of any *Listeria* spp. in a product be

investigated as it could indicate a failure in procurement, preparation and/or storage of food materials²⁴.

The prevalence of *L. monocytogenes* observed in cold-sliced meats in the UK in the present study (3.7%) was lower than that found in Belgium (7.5%)³⁰, Denmark (5%)³¹, Norway (11%)³² but higher than that reported in the US (3.5%)³³ and previously in the UK (2.0%)³⁴. The prevalence of *L. monocytogenes* found in cold-sliced meats at the end of shelf-life in the current study (4.2%; 1.0% >100 cfu/g) concurred with that found previously in sliced meats examined at the end of shelf-life during 2003 in the UK (4.8%; 0.9% >100 cfu/g)³⁵. This study has also highlighted contributory factors likely to cause problems with retail sliced meats. Sliced meats were more frequently contaminated with both *Listeria* spp. and *L. monocytogenes* when they were from premises that sliced meats to order, sold in large pack sizes, and stored or displayed above 8°C. Continuous efforts should be made to improve food safety of sliced meats by adequate cooking and appropriate hygienic measures to avoid contamination, and through use of appropriate refrigeration temperatures and shelf-life to prevent growth of *L. monocytogenes* in these products. Storage of chilled foods such as meats must comply with Regulation (EC) No 853/2004 on the hygiene of foodstuffs¹¹, i.e. should not be kept at temperatures that might result in a risk to health.

Management food hygiene training and the presence of hazard analysis systems in food premises has been shown to make a significant contribution to an improvement in the microbiological quality of ready-to-eat foods³⁶. This is corroborated by the results from this study. The implementation of a hazard analysis system or similar food safety management plans in food premises provides a pragmatic framework for good hygiene practice. Compliance with the principles of HACCP and associated relevant supervision and instruction and/or food hygiene training for all employees is a legal requirement¹¹. Evidence from this study also highlights smaller retail premises (as indicated by Consumer at Risk scores), and premises with poor Confidence in Management scores, as an area for concern with regard to microbiological quality of ready-to-eat food (including presence of *L. monocytogenes*) and food safety risks.

Several European countries, including the UK, are experiencing an increased incidence of listeriosis among persons in their 'golden years' (≥60 years of age)³⁻⁶. The serogroups most often causing human infection in the UK are serogroups 4b and 1/2a, with the subtype 4b AFLP I being most common in England and Wales^{6,37}. A preliminary case/case study of human *L. monocytogenes* subtypes in relation to food exposure history from 2005 to 2007 in England and Wales found that those infected with serotype 1/2a were more likely to report the consumption of tongue, chicken pies, various fish, cheese, sandwiches and salads, and those infected with 4b with consumption of cold cooked beef and sandwiches containing hard cheese⁶. The same study further revealed associations

between exposures of *L. monocytogenes* sero-AFLP types, to pork or dairy products (4b I), fish and dairy products (1/2a IX), and sliced meats (ham, chicken turkey), cheese, and sandwiches (1/2a VII)⁶. The predominant serogroup of *L. monocytogenes* recovered in this study from sliced meats (ham, beef, chicken, tongue) and sandwich (egg, ham, tuna, cheese with & without salad) isolates in the present study was serotype 1/2a (63%), with AFLP subtypes IX and VII prevalent. However, 21% of meat and sandwich food isolates were serotype 4b, with AFLP subtypes I and IV most common. The predominant serogroup recovered from all food isolates in the UK from 2005 to 2007 was also serogroup 1/2a (52%), of which 80% were AFLP IX (60%) or VII (20%) (HPA unpublished information).

Consumers are believed to benefit from clear recommendations on good food hygiene practice (i.e. at what temperature to keep food chilled at all times), and from being encouraged to take careful note of the shelf-life of food in their refrigerators³. Packaging/labelling safety based guidance on the proper storage and handling of refrigerated ready-to-eat foods should therefore help to reduce the risk of listeriosis. A small proportion of ready-to-eat foods in this study had this information present in a format that many consumers would have difficulty in reading and should be avoided, i.e. using 6 point font size (shown here for example). Labelling design should continue to make every effort on legibility and clarity, particularly as there is a growing elderly population where there will be more people with sub-optimal vision³⁸. This is being currently considered as part of an EU food labelling review³⁸.

Dietary recommendations about when to avoid certain foods and educational messages about food preparation are also important. Advice from the Food Standards Agency to pregnant women in the UK warns against consumption of pâté, soft mould-ripened cheeses such as camembert and brie, and blue veined cheeses, and to cook raw meat thoroughly³⁹. Dietary advice on the avoidance of high-risk foods should also be provided routinely to other susceptible groups, such as the elderly population. An ACMSF Group⁴⁰ is currently considering this issue and is to provide recommendations to the Food Standards Agency on whether current advice needs to be amended.

Listeriosis is predominantly transmitted by the consumption of contaminated ready-to-eat foods³, therefore effective responses are essential to control this pathogen in the food chain. Significant progress has been made in recognising foods which present a risk of *L. monocytogenes* infection, and in developing strategies and processes that can minimise these risks. Current EC microbiological criteria indicate that levels of *L. monocytogenes* at ≤ 100 cfu/g in ready-to-eat foods within shelf-life are legally satisfactory⁹. This study highlights that further emphasis must be given in the reduction of *L. monocytogenes* in high-risk foods, such as sliced meats and sandwiches, which are consumed without any further treatment.

Acknowledgements

The authors would like to thank all the staff in the Environmental Health Departments throughout the UK who collected samples for this study, and all the staff in HPA, HPA Collaborating and other Official Food Control laboratories who performed the microbiological examinations. Thanks are extended to FSML (HPA Centre for Infections) for characterising *Listeria monocytogenes* isolates, to David Lock at LACORS for co-ordinating the participation of Environmental Health Officers and advice from the LACORS Food Examination Focus Group and Food Hygiene Focus Group, to the HPA Regional FWE Coordinators Forum for their advice in preparing the sampling protocols, and to Lillian Hucklesby for co-ordinating data entry and validation.

References

1. Adak, G.K., Long, S.M., O'Brien, S.J. Trends in indigenous foodborne disease and deaths, England and Wales: 1992 to 2000. *Gut* 2002; **51**: 832-41.
2. Gillespie, I.A., McLauchlin, J., Grant, K.A., Little, C.L., Mithani, V., Penman, C. and Regan, M. Changing pattern of human listeriosis in England and Wales, 2001-2004. *Emerg. Inf. Dis.* 2006; **12**: 1361-6.
3. European Food Safety Authority (EFSA). Scientific Opinion of the Panel on Biological Hazards on a request from the European Commission on Request for updating the former SCVPH Opinion on *Listeria monocytogenes* in ready-to-eat foods and the related risk for human illness. *The EFSA Journal* 2007; **599**: 1-42.
4. Goulet, V., Hedberg, C., Le Monnier, A., de Valk, H. Increasing incidence of listeriosis in France and other European Countries. *Emerg. Inf. Dis.* 2008; **14**: 734-40.
5. Denny, J., McLauchlin, J. Human *Listeria monocytogenes* infections in Europe - an opportunity for improved European surveillance. *Euro Surveill* 2008; **13** (13). Available at: http://www.eurosurveillance.org/edition/v13n13/080327_5.asp. Accessed 14 May 2008.
6. Gillespie, I., McLauchlin J. Update on listeriosis in England and Wales, December 2007. Advisory Committee on the Microbiological Safety of Food, Paper ACM/879. Available at: <http://www.food.gov.uk/multimedia/pdfs/committee/879listeria.pdf>. Accessed 7 May 2008.
7. Rocourt J, BenEmbarak P, Toyofuku H, Schlundt J. Quantitative risk assessment of *Listeria monocytogenes* in ready-to-eat foods: the FAO/WHO approach. *FEMS Immunol. Med. Microbiol* 2003; **35**: 263-7.
8. Bell C, Kyriakides A. *Listeria*. A practical approach to the organism and its control in foods, 2nd Ed. Blackwell Publishing: Oxford, 2005.
9. European Commission (EC). Regulation (EC) No 2073/2005 of 15 November 2005 on microbiological criteria for foodstuffs. *Off J Europ Union* 2005; **L338**: 1-26.
10. European Commission (EC). Opinion of the Scientific Committee on Veterinary Measures relating to Public Health on *Listeria monocytogenes*, 23 September 1999. Available at: http://ec.europa.eu/food/fs/sc/scv/out25_en.pdf. Accessed on 7 May 2008.
11. European Commission (EC). (2004). Regulation (EC) No 852/2004 of the European Parliament and of the Council of 29 April 2004 on the hygiene of foodstuffs. *Off J Europ Union* 2004; **L139**: 1-54.
12. Meldrum R, Ribeiro CD, Simmons MD, Worthington D, Griffith C. The Welsh Food Microbiological Forum and the all-Wales shopping basket sampling program: A

- model for the surveillance of microbiological quality in ready-to-eat foods. *J Env Health* 2003; **65**: 24-8.
13. Willis C, Greenwood M. Wessex shopping basket survey – a structured approach to local food sampling. *Int J Environ Health Res* 2003; **13**: 349-59.
 14. Food Standards Agency (FSA). Food Law Code of Practice. London. FSA, 2006.
 15. Local Authorities Co-ordinators of Regulatory Services (LACORS). LACORS Guidance Food Sampling for Microbiological Examination, Issue 2, 2006. Available at: <http://www.lacors.com>. Accessed 7 May 2008.
 16. Health Protection Agency. Direct enumeration of *Escherichia coli*. National Standard Method Standard F20 Issue 1, 2004. Available at: <http://www.hpa-standardmethods.org.uk/documents/food/pdf/F20.pdf>. Accessed 7 May 2008.
 17. Health Protection Agency. Enumeration of β -glucuronidase positive *Escherichia coli* – Most Probable Number method. National Standard Method: F22 Issue 1, 2005. Available at: <http://www.hpa-standardmethods.org.uk/documents/food/pdf/F22.pdf>. Accessed 7 May 2008.
 18. Health Protection Agency. Enumeration of *Staphylococcus aureus*. National Standard Method: F12 Issue 1, 2005. Available at: <http://www.hpa-standardmethods.org.uk/documents/food/pdf/F12.pdf>. Accessed 7 May 2008.
 19. Health Protection Agency. Detection and enumeration of, *Listeria monocytogenes* and other *Listeria* species. National Standard Method F19 Issue 2, 2007. Available at: <http://www.hpa-standardmethods.org.uk/documents/food/pdf/F19.pdf>. Accessed 7 May 2008.
 20. Doumith M, Buchrieser C, Glaser P, Jacquet C, Martin P. Differentiation of the major *Listeria monocytogenes* serovars by multiplex PCR. *J. Clin. Microbiol.* 2004; **42**: 3819-22.
 21. Guerra MM, Bernardo F, McLauchlin J. Amplified fragment length polymorphism (AFLP) analysis of *Listeria monocytogenes*. *Syst. Appl. Microbiol.* 2002; **25**: 456-61.
 22. Gilbert, R.J., de Louvois, J., Donovan, T., Little, C., Nye, K., Riberio, C.D., Richards, J., Roberts, D. and Bolton, F.J. Guidelines for the microbiological quality of some ready-to-eat foods sampled at the point of sale. *Comm. Dis. Pub. Health* 2000; **3**: 163-167.
 23. Anon. The Food Hygiene (England) Regulations 2006 (SI 2006/14). Available at: www.opsi.gov.uk/si/si2006/ukSI_20060014_en.pdf. Accessed 9 July 2008.
 24. CFA. Position Statements, Temperature. Available at: <http://www.chilledfood.org/MEDIA/POSITION+STATEMENTS/temperature.htm>. Accessed 9 July 2008.
 25. British Sandwich Association Manufacturer Code of Practice, Revised August 2007. Available at: http://www.sandwichesonline.org.uk/about_the_bsa/manufacture_code_of_practice.htm. Accessed 7 May 2008.
 26. Meldrum, R., and R.M.M. Smith. 2007. Occurrence of *Listeria monocytogenes* in sandwiches available to hospital patients in Wales, UK. *J. Food Prot.* 70: 1958-1960.
 27. Wilson I.G. Occurrence of *Listeria* species in prepacked retail sandwiches. *Epidemiol Infect* 1996; **117**: 89-93.
 28. Food Safety Authority of Ireland (FSAI). Microbiological safety of pre-packed sandwiches, 2002. Available at: http://www.fsai.ie/surveillance/food_safety/microbiological/3rdQuarter_prepacked_sandwiches.pdf. Accessed 12 May 2008.
 29. Little, C.L., Barrett, N.J., Grant, K., McLauchlin, J. (2008) Microbiological safety of sandwiches from hospitals and other healthcare establishments in the UK with a focus on *Listeria monocytogenes* and other *Listeria* spp. *Journal of Food Protection*, 71: 309-318.
 30. Uyttendaele, M., de Troy, P. and Debevere, J. (1999) Incidence of *Listeria monocytogenes* in different types of meat products on the Belgian retail market. *International Journal of Food Microbiology* 53, 75–80.

31. Norrung, B., Andersen, J.K. and Schlundt, J. (1999) Incidence and control of *Listeria monocytogenes* in foods in Denmark. *International Journal of Food Microbiology* 53, 195–203.
32. Rovik, L.M. and Yndestad, M. (1991) *Listeria monocytogenes* in foods in Norway. *International Journal of Food Microbiology* 13, 97–104.
33. Levine, P., Rose, B., Green, S., Ransom, G. and Hill, W. (2001) Pathogen testing of ready-to-eat meat and poultry products collected at federally inspected establishments in the United States, 1990 to 1999. *Journal of Food Protection* 64, 1188–1193.
34. Elson, R., Burgess, F., Little, C.L., Mitchell, R.T. (2004). Microbiological examination of ready-to-eat cold sliced meats and pâté from catering and retail premises in the United Kingdom. *Journal of Applied Microbiology*, 96: 499-509.
35. Sagoo, S.K., Little, C.L., Allen, G., Williamson, K., Grant, K.A. (2007). Microbiological safety of retail vacuum packed and modified atmosphere packaged cooked meats at end of shelf-life. *Journal of Food Protection*, 70: 943-951.
36. Little, C.L., Lock D., Barnes J., Mitchell, R.T. The microbiological quality of food in relation to hazard analysis systems and food hygiene training in UK catering and retail premises. *Communicable Disease & Public Health*, 2003; 6: 250-258.
37. McLauchlin, J. The pathogenicity of *Listeria monocytogenes*: A public health perspective. *Rev. Med. Microbiol.* 1997; 8: 1-14.
38. Food Standards Agency, 2008. Food labelling. Clear Food Labelling Guidance, Revised 2008. Available at: <http://www.food.gov.uk/news/newsarchive/2008/jun/revlabguid>. Accessed 5 June 2008.
39. Food Standards Agency, 2008. Eat Well, Be Well. When you're pregnant – what to avoid. Available at: <http://www.eatwell.gov.uk/agesandstages/pregnancy/whenyrpregnant/#cat226049>. Accessed 5 June 2008.
40. Advisory Committee on the Microbiological Safety of Food. ACMSF minutes: 6 December 2007. Available at: <http://acmsf.food.gov.uk/acmsfmeets/acmsf2007/acmsf061207/acmsfmin061207>. Accessed 12 May 2008.

Annex I: Participating Laboratories and Local Authority Food Liaison Groups and number of samples

Table 1a. Participating HPA and HPA Collaborating Laboratories and number of samples

| HPA Region | HPA/HPA Collaborating Laboratory | Number of samples |
|---|----------------------------------|-------------------|
| East | Chelmsford | 377 |
| | Norwich | 188 |
| East Midlands | Leicester | 86 |
| | Lincoln | 316 |
| London | London FWEM ¹ | 176 |
| South East | Ashford | 344 |
| | Brighton | 379 |
| | WEMS ² | 1094 |
| North East and Yorkshire and the Humber | Leeds | 111 |
| | Newcastle | 451 |
| | Hull | 285 |
| | Sheffield | 241 |
| North West | Carlisle | 64 |
| | Chester | 230 |
| | Preston | 993 |
| South West | Bristol | 244 |
| | Exeter | 275 |
| | Gloucester | 254 |
| | Plymouth | 40 |
| | Truro | 89 |
| West Midlands | Birmingham | 149 |
| | Coventry | 37 |
| | Shrewsbury | 169 |
| | Hereford | 93 |
| Total | | 6685 |

1, London Food, Water & Environmental Microbiology Services Laboratory

2, Wessex Environmental Microbiology Services

Table 1b. Other participating Official Food Control Laboratories in Wales, Scotland, Northern Ireland & England and number of samples examined.

| Country | Laboratory | Number of samples |
|--------------|--|-------------------|
| Wales | NPHS-W Microbiology Cardiff | 48 |
| | NPHS-W Microbiology Rhyl | 37 |
| Ireland | Belfast City Hospital | 72 |
| Scotland | Aberdeen City Council Public Analysts | 45 |
| | Edinburgh Analytical and Scientific Services | 46 |
| | Glasgow Scientific Services | 48 |
| England | Kings Lynn & West Norfolk | 2 |
| Total | | 298 |

Table III: Participating Food Safety Liaison Groups and number of samples

| Local Authority Food Liaison Group | Number of Samples |
|--|--------------------------|
| Berkshire | 156 |
| Buckinghamshire | 15 |
| Cambridgeshire | 125 |
| Cheshire | 115 |
| Cornwall | 89 |
| Cumbria | 110 |
| Derbyshire | 185 |
| Devon | 206 |
| Dorset | 27 |
| Durham | 149 |
| East Sussex | 113 |
| Essex | 227 |
| Gloucester | 254 |
| LFCG ¹ Greater London NE Sector | 12 |
| LFCG Greater London NW Sector | 30 |
| LFCG Greater London SE Sector | 9 |
| LFCG Greater London SW Sector | 45 |
| Greater Manchester | 421 |
| Hampshire & Isle Of Wight | 541 |
| Hereford & Worcester | 135 |
| Hertfordshire & Bedfordshire | 73 |
| Humberside | 285 |
| Kent | 594 |
| Lancashire | 525 |
| Leicestershire | 86 |
| Lincolnshire | 143 |
| Merseyside | 104 |
| North Yorkshire | 105 |
| Northamptonshire | 26 |
| Northern Ireland Food Group ² | 72 |
| Norfolk | 148 |
| Nottinghamshire | 124 |
| Northumberland | 38 |
| Oxfordshire | 218 |
| Scottish Food Enforcement Liaison Committee ³ | 141 |
| Shropshire | 78 |
| Somerset | 108 |
| South West Yorkshire | 198 |
| Staffordshire | 1 |
| Suffolk | 66 |
| Surrey | 189 |
| Tees Valley | 115 |
| Tyne & Wear | 61 |
| Wales South East Group | 49 |
| Wales North Group | 47 |
| Wales West Group | 35 |
| Warwickshire | 1 |
| West Midlands | 173 |
| West of England | 115 |
| West Sussex | 207 |
| Wiltshire | 145 |
| Total | 6983 |

1, London Food Co-ordinating Group; 2, Northern Ireland Food Group consists of Eastern, Northern, Southern & Western Groups; 3, SFELG consists of Central Scotland, Fife & Tayside, Lothian & Scottish Borders, North Scotland, and West of Scotland

Annex II Sampling Questionnaire



STUDY 28: LACORS/HPA CO-ORDINATED FOOD LIASION GROUP STUDY: PROFORMA

Focused Shopping Basket Sampling of selected foods from retail premises with a
focus on *Listeria monocytogenes* and *Listeria* spp.

Add Laboratory Name & Address

Local Authority Sample Reference Number(s).....

Laboratory Sample Number(s).....

LACORS/HPA Shopping Basket; May 2006 – April 2007
LABORATORY NAME.....

Local Authority Sampling details:

- | | |
|--|--|
| 1. Local Authority..... | 2. Food Liaison group..... |
| 3. Samples collected by..... (EHO/EHT/ Sampling Officer) | 4. Sample collected at..... (time) on (date)/...../..... |
| 5. LA Premises Reference Number..... | 6. LA Sample Reference Number(s)..... |

Premises details:

7. Name of premises.....
8. Address..... Postcode.....
9. Type of premises
- | | | | | | |
|--------------------------|--------------------------|------------------|--------------------------|-------------------|--------------------------|
| Supermarket pre-packed | <input type="checkbox"/> | Delicatessen | <input type="checkbox"/> | Market stall/shop | <input type="checkbox"/> |
| Supermarket delicatessen | <input type="checkbox"/> | Butcher | <input type="checkbox"/> | Bakery | <input type="checkbox"/> |
| Sandwich bar | <input type="checkbox"/> | Convenience shop | <input type="checkbox"/> | Farmer's Market | <input type="checkbox"/> |
| Continental market | <input type="checkbox"/> | Other | <input type="checkbox"/> | (Specify)..... | |
- 10 (A) Inspection Rating Category (A - E): (B) Consumers at risk score (0 - 15):
- (c) Confidence in Management/Control Systems Score (0 - 30).....
- 11 (A). Does the premises comply with the principles of HACCP (Article 5 of EC 852/2004)? YES ☐ NO ☐
- 11 (B). Has the manager undertaken food hygiene training? YES ☐ NO ☐; If YES, what level? Foundation ☐ Intermediate ☐ Advanced ☐ Other ☐ (specify).....

12. Sample details:

- A. Sliced Meat *** ☐ & Specify type: Ham ☐ Chicken ☐ Beef ☐ Tongue ☐
- * For Prepacked sliced meat samples only : Please take duplicate samples of the same batch for testing within and at end of shelf-life*
- B. Sandwiches** ☐ & Specify type: Ham ☐ Cheese ☐ Tuna ☐ Egg mayo ☐
- & does the sandwich contain salad or a salad component i.e. tomato, lettuce, cucumber? YES ☐ NO ☐
- C. Hard Cheese** ☐ & Specify type: Cheddar ☐ Gloucester ☐ Lancashire ☐ Leicester ☐
- & is the cheese made from: Unpasteurised milk ☐ or Pasteurised milk ☐
- D. Spreadable cheese** ☐ & Specify type: Cheese spread ☐ Cream cheese ☐
- & is it: Plain ☐ Or Flavoured ☐ (specify flavour).....
- E. Butter** ☐ & Specify type: Salted ☐ Lightly salted (<1%) ☐ Unsalted ☐ Unknown ☐
- & is the butter made from: Unpasteurised milk ☐ or Pasteurised milk ☐
- F. Probiotic drinks** ☐ & Specify type: Plain ☐ or Flavoured ☐ (specify flavour).....
- G. Confectionery Product** ☐ containing Fresh Cream ☐ Or Synthetic/ imitation cream ☐

13. Is the product cooked on site: YES ☐ NO ☐ Not Applicable ☐
14. Is the product: Prepacked ☐ **Go to Q 15** or Cut/ Made to order ☐ **Go to Q 17**
15. Is the product: Vacuum packed ☐ Modified atmosphere packed ☐ or Packed in normal atmosphere ☐
16. Is the product packed on the premises: YES ☐ NO ☐
17. What is the pack size: <100g ☐ 100-<200g ☐ 200- <300g ☐ ≥300g ☐ (specify).....
18. Was the product displayed/stored at: Equal/below 8°C ☐ & Specify temperature.....°C
or above 8°C ☐ & Specify temperature.....°C

Labelling and Storage, Shelf-life and data marking details:

19. Specify Product Name **exactly** as it appears on the label/container:.....
20. Brand/ Producer (specify):..... Not applicable ☐
21. What is the storage temperature information on the packaging? Keep refrigerated ☐
Particular temperature given ☐ & specify.....°C
No temperature indicated ☐
22. What are the shelf-life instructions on the packaging on opening? Consume within 3 days ☐
Particular period given ☐ & specify.....
None ☐
23. Product Date (specify): Best before Date...../...../..... Use By Date...../...../..... Display Date...../...../..... Not Known ☐
24. Is the storage, shelf life, date marking information: Easily visible & clearly legible YES ☐ NO ☐
In black type on a white background YES ☐ NO ☐ If NO Specify the colour of the type & Background.....
25. What is the font size of the type/print used for this information: 10 point ☐ 8 point ☐ Other ☐ specify
26. Is the product labelled as organic? YES ☐ NO ☐
27. What is the country of origin?.....
28. Is there a batch code? YES ☐ NO ☐ If YES please specify code.....
29. Is there an EU approval number? YES ☐ NO ☐ If YES please specify code.....

LACORS/HPA Shopping Basket; May 2006 – April 2007
LABORATORY NAME.....

Laboratory details:

Sample(s) received in laboratory..... (time) on (date)/...../.....
 Sample(s) received by.....
 Sample(s) received from.....
 Temperature on receipt°C
 Within the cool box is a temperature monitoring device used e.g. data logger : YES ☐ NO ☐
 Sample(s) examined. (time) on (date)/...../.....

RESULTS

Recording results

Please record the results of count/g tests as **ACTUAL NUMBERS** in the appropriate box within the table. Only place ticks in the column headed (<10/<20), i.e. the limit of detection for that test, and columns headed ND and Detected.

TABLE 1. Samples Tested on Receipt at Laboratory

Laboratory Sample No.....

| | ND | Detected | <10/<20 | 20-<10 ² | 10 ² -<10 ³ | 10 ³ -<10 ⁴ | 10 ⁴ -<10 ⁵ | 10 ⁵ -<10 ⁶ | 10 ⁶ -<10 ⁷ | ≥10 ⁷ |
|---|----|----------|---------|---------------------|-----------------------------------|-----------------------------------|-----------------------------------|-----------------------------------|-----------------------------------|------------------|
| <i>E.coli</i> /g | | | | | | | | | | |
| <i>Staphylococcus aureus</i> /g | | | | | | | | | | |
| <i>Listeria</i> spp. (total) / 25g or g | | | | | | | | | | |
| <i>L.monocytogenes</i> / 25g or g | | | | | | | | | | |

Microbiological Quality: Satisfactory ☐ Acceptable ☐ Unsatisfactory ☐ Unacceptable/Potentially hazardous ☐

TABLE 2. Sliced Meat Samples Tested at the End of Shelf-life

Storage temperature

Were the samples stored in a 6±2°C incubator? YES ☐ NO ☐

If NO please record the temperature of the monitored sample refrigerator that samples were stored in:.....°C

Laboratory Sample No.....

| | ND | Detected | <10/<20 | 20-<10 ² | 10 ² -<10 ³ | 10 ³ -<10 ⁴ | 10 ⁴ -<10 ⁵ | 10 ⁵ -<10 ⁶ | 10 ⁶ -<10 ⁷ | ≥10 ⁷ |
|---|----|----------|---------|---------------------|-----------------------------------|-----------------------------------|-----------------------------------|-----------------------------------|-----------------------------------|------------------|
| <i>E.coli</i> /g | | | | | | | | | | |
| <i>Staphylococcus aureus</i> /g | | | | | | | | | | |
| <i>Listeria</i> spp. (total) / 25g or g | | | | | | | | | | |
| <i>L.monocytogenes</i> / 25g or g | | | | | | | | | | |

Microbiological Quality: Satisfactory ☐ Acceptable ☐ Unsatisfactory ☐ Unacceptable/Potentially hazardous ☐

Date *L. monocytogenes* isolates sent to the Food Safety Microbiology Laboratory, HPA Centre for Infections.....

Date *S.aureus* isolates sent to the Food Safety Microbiology Laboratory, HPA Centre for Infections.....

MICROBIOLOGISTS COMMENTS.....

Further Report:

Identification and type:

Signature Date reported

**WHERE POSSIBLE THE LABORATORY SHOULD MAKE AND RETAIN A PHOTOCOPY OF THE
 PRODUCT LABEL FOR FUTURE REFERENCE**

Methods as defined in LACORS/HPA Shopping Basket Study (Study 28); Annex 6

COPY TO: Dr Satnam Sagoo, Environmental & Enteric Diseases Department, HPA Centre for Infections,
 61 Colindale Avenue, London NW9 5EQ

Page 3 of 3