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ADVISORY COMMITTEE ON THE MICROBIOLOGICAL SAFETY OF FOOD

INFORMATION PAPER

**Microbiological Examination of Dried Spices and Herbs from Production
and Retail Premises in the United Kingdom**

The final report of this study carried out in 2004 is attached for Members' information.

**Secretariat
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Microbiological Examination of Dried Spices and Herbs from Production and Retail Premises in the United Kingdom

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On behalf of the Food standards Agency, Local Authorities Co-ordinators of Regulatory Services and the Health Protection Agency

Summary

As part of the European Commission co-ordinated programme for the official control of foodstuffs, a study of dried spices and herbs from retail and production premises was undertaken in the UK during 2004 to determine the microbiological status of these products. According to microbiological criteria in Recommendation 2004/24/EC and European Spice Association specifications, 96% of 2833 retail samples and 92% of 132 production batches were of satisfactory/acceptable quality. *Salmonella* spp. was detected in both 1% of dried spices and herbs sampled at retail and production. Overall, a small proportion of herbs and spices contained high counts of *B. cereus* (1%, $\geq 10^4$ cfu/g), *C. perfringens* (0.4%, $\geq 10^3$ cfu/g) and *E. coli* (2.1%, $\geq 10^2$ cfu/g). Ninety percent of samples examined were recorded as being 'ready-to-use'. The potential public health risk of using spices and herbs as an addition to ready-to-eat foods that potentially undergo no further processing is therefore highlighted in this study. Prevention of microbial contamination in dried herbs and spices lies in the application of good hygiene practices during growing, harvesting and processing from farm to fork, and effective decontamination. In addition, the importance of correct food handling practices and usage of herbs and spices by end users cannot be overemphasised.

Introduction

Spices and herbs are valued for their distinctive flavours, colours and aromas and are among the most versatile and widely used ingredient in food preparation and processing throughout the world. Spices are generally grown in the tropical and sub tropical regions, whereas herbs are grown in the more temperate parts of the world. As with many other agricultural products, spices and herbs may be exposed to a wide range of microbial contamination during pre- and post-harvest, including processing, storage, distribution, sale and/or use¹⁻³. For example, the traditional method of drying spices and herbs post harvest is to spread them out on the ground to dry under the sun; however this potentially exposes them to the risk of contamination. The use of enclosed drying systems can reduce this problem considerably.

Dried spices and herbs may therefore contain high levels of microbial contamination, depending on whether they have received a form of treatment or not^{3,4}. Bacteria of public health importance, such as *Salmonella*, may be present and this could potentially create a public health risk depending on the end use, such as addition to ready-to-eat foods that undergo no further processing. Some outbreaks of salmonellosis have been linked to the consumption of foods seasoned with black or white pepper^{5,6}, chilli, turmeric powder⁷, paprika powder flavoured potato chips⁸, and aniseed containing tea⁹. *Bacillus cereus* may also be present in spices and herbs, usually at counts below 10^3 cfu/g but may multiply to high levels ($10^5 - 10^6$ cfu/g) in food to which it is added; this may be sufficient to cause food poisoning if the food is inappropriately handled or stored^{4,6-7}.

The control of microbial contamination in these products lies in the application of good hygiene practices in the production/harvesting area, processing, and personnel. In addition to good hygienic practices, many spice and herb suppliers apply control processes based on steam or dry heat treatments. These processes are designed to minimise the risk from pathogens and to retain the natural characteristics of the products. Codex has produced a Code of Hygienic Practice for Spices and Dried Aromatic Plants¹⁰ for spice and herb

growers/suppliers to help minimize microbial food safety hazards. There are no microbiological standards for dried spices and herbs in EC legislation, however the Codex Code of Hygienic Practice¹⁰ specifies that dried spices and herbs should be free from pathogenic microorganisms at levels that may represent a hazard to health and further requires that *Salmonella* should be absent in treated ready-to-eat spices. The European Spice Association (ESA) also specifies that *Salmonella* should be absent in 25g of spice, *Escherichia coli* to be present at less than 10² cfu/g, and other bacteria requirements to be agreed between buyer and seller^{11,12}. To gain insight on the prevalence of *Salmonella*, and the frequency and levels of *Bacillus cereus*, *Clostridium perfringens* and *E. coli* contamination in these spice and herb products from production and retail premises, the European Commission Recommendation 2004/24/EC required Member States to carry out a co-ordinated programme of sampling and testing of dried spices and herbs¹³. Reported here is a detailed analysis of the UK contribution to the 2004 EC co-ordinated food sampling programme.

Materials and Methods

Sample collection

Dried spices and herbs collected from retail and production premises were examined by 38 Official Food Control laboratories (Health Protection Agency (HPA), HPA Collaborating Laboratories, National Public Health Service (NPHS)-Wales and Public Analysts) in the UK between 1 July and 31 December 2004 according to a standardised protocol. Five sample units (5 x ≥30g) were collected from each batch at production premises using class attribute sampling plans as provided in Commission Recommendation 2004/24/EEC¹³. At retail premises single samples (≥30g) were collected. Samples were collected and transported to laboratories by staff from 293 local Environmental Health Departments, involving 271 Local Authority Food Liaison Groups (Annex 2), in accordance with the FSA Food Law Code of Practice¹⁴ and LACORS guidance on microbiological food sampling¹⁵.

Information on samples and premises was obtained by observation and enquiry and recorded on a standard proforma. Additional information collected included the type of spice or herb, country of origin, packing details, processing and treatment details, existence of a hazard analysis system and the level of food hygiene training received by the manager. Food hygiene inspections are carried out in a way that focuses enforcement authority resources on premises presenting most risk to consumers. To do this, food hygiene inspections are carried out in accordance with FSA Food Law Code of Practice¹⁴ which specifies that, amongst other factors, the number of consumers at risk and confidence in management control systems (including the application of HACCP based systems) should be assessed to produce a risk rating of the premises. The risk rating determines the frequency of inspection and at the time of this study ranged from Category A (highest risk, inspected every 6 months) to F (lowest risk, inspected every 5 years).

Sample examination

Salmonella spp., *E. coli*, Enterobacteriaceae, *B. cereus* and other *Bacillus* spp. and *C. perfringens* were detected or enumerated in accordance with HPA Standard Microbiological Methods¹⁶⁻²⁰. Isolates of *Salmonella* spp. were sent to the Laboratory of Enteric Pathogens (LEP), Health Protection Agency Centre for infection (HPA CfI), for typing and further characterisation. Isolates of *B. cereus* and other *Bacillus* spp. at $\pm 10^5$ cfu/g were sent to the Food Safety Microbiology Laboratory (FSML) HPA CfI for further confirmation.

The microbiological status of dried herb and spice samples were assessed using criteria in Recommendations 2004/24/EC¹³ and the European Spice Association (ESA) specifications¹² (Tables 1 and 2).

Table 1: Classification of dried herbs and spices from production premises as recommended by microbiological criteria within Recommendation 2004/24/EC¹³ and ESA¹²

Microorganism	Microbiological quality*
<i>Bacillus cereus</i>	n=5, c=1, m= 10 ³ cfu/g, M =10 ⁴ cfu/g
<i>Clostridium perfringens</i>	n=5, c=1, m= 10 ² cfu/g, M =10 ³ cfu/g
<i>Salmonella</i> spp.	n = 5, c = 0, Absent in 25 g
<i>Escherichia coli</i>	n = 5, c=1, m= 10 cfu/g, M = 10 ² cfu/g

*Parameters defined as follows:

n = number of units comprising the sample

m = limit below which all results are considered satisfactory

M = limit above which all results are considered unsatisfactory

c = number of sample units that may fall between m and M

For *B. cereus* and *C. Perfringens* the status of a batch is considered to be:

- Satisfactory where all the values are m or less
- Acceptable where the maximum of c values are between m and M
- Unsatisfactory if one or more values is/are equal or above M or more than c values between m and M

For *Salmonella* spp. the status of a batch is considered to be:

- Satisfactory where all the values are not detected in 25g
- Unsatisfactory where one or more values are detected in 25g

Table 2: Classification of dried herbs and spices from retail premises as recommended by microbiological criteria within Recommendation 2004/24/EC¹³ and ESA¹²

Microorganism	Microbiological quality (cfu per gram unless stated)		
	Satisfactory	Acceptable	Unsatisfactory
<i>Bacillus cereus</i>	<10 ³	10 ³ -<10 ⁴	≥10 ⁴
<i>Clostridium perfringens</i>	<10 ²	10 ² -<10 ³	≥10 ³
<i>Salmonella</i> spp.	*ND in 25g	–	Detected in 25g
<i>Escherichia coli</i>	<10	10-<10 ²	≥10 ²

*ND = not detected in 25g

For the purposes of the EC co-ordinated sampling programme, the Enterobacteriaceae count was used as an indicator for possible irradiation or other treatments of spices and herbs (Recommendation 2004/24/EC)¹³. Samples that had an Enterobacteriaceae count of ≤100 cfu/g were therefore considered as having been irradiated or submitted to heat treatments¹³.

Statistical Analysis

Descriptive and statistical analysis of the data was undertaken using Microsoft Excel and Epi Info version 6.04d. Relative proportions were compared using the chi-squared (χ^2) and Fisher's exact tests. A probability value of less than 5% was deemed to be significant.

Results

Microbiological status of dried spices and herbs

In total 132 (660 sample units) batches and 2833 samples of dried spices and herbs were tested from production and retail establishments, respectively. Of these, 2199 (74%) were spices (*Capsicum* spp., *Piper* spp. and other types) and 766 (26%) were herbs.

Production Establishments

Applying the criteria in Recommendation 2004/12/EC and ESA specifications (Table 1), 75% (98/132) of the batches of dried herbs and spices were of satisfactory microbiological quality, 17% (23) were of acceptable quality, and a further 8% (11) were of unsatisfactory quality due to the presence of *Salmonella* spp., *E. coli* (ranging from 2.4×10^3 – 1.0×10^7 cfu/g) and/or high levels of *B. cereus* (ranging from 1.0×10^5 – 2.3×10^7 cfu/g) (Table 3).

Table 3. Microbiological status of dried spices and herbs according to Recommendation 2004/24/EC¹³ and ESA¹² specifications from production premises in the UK

Microorganism	Product Identification	Number of samples	Analysis results		
			Satisfactory	Acceptable	Unsatisfactory
<i>Salmonella</i> spp.	<i>Capsicum</i> spp.	18	18	0	0
	<i>Piper</i> spp.	28	28	0	0
	Other spices	63	61	0	2
	Herbs	23	23	0	0
<i>Bacillus cereus</i>	<i>Capsicum</i> spp.	18	16	2	0
	<i>Piper</i> spp.	28	22	4	2
	Other spices	63	54	6	3
	Herbs	23	16	5	2
<i>Clostridium perfringens</i>	<i>Capsicum</i> spp.	18	17	1	0
	<i>Piper</i> spp.	28	27	1	0
	Other spices	63	61	2	0
	Herbs	23	22	1	0
<i>Escherichia coli</i>	<i>Capsicum</i> spp.	18	15	2	1
	<i>Piper</i> spp.	28	27	1	0
	Other spices	63	62	1	0
	Herbs	23	22	0	1
Enterobacteriaceae			Analysis results		
				£100 cfu/g	>100 cfu/g
	<i>Capsicum</i> spp.	18		7	11
	<i>Piper</i> spp.	28		19	9
	Other spices	63		25	38
	Herbs	23		8	15

Retail Premises

Applying the criteria in Recommendation 2004/12/EC¹³ and ESA specifications¹² (Table 2), 85% (2415/2833) of the dried herbs and spices samples were of satisfactory microbiological quality and 11% (307) were of acceptable quality. A further 4% (111) were of unsatisfactory quality due to the presence of *Salmonella* spp., and/or high levels of *E. coli* (ranging from 1.0×10^3 - $\pm 10^7$ cfu/g), *C. perfringens* (ranging from 1.0×10^3 – 2.4×10^5 cfu/g) and *B. cereus* (ranging from 1.7×10^5 – 1.2×10^6 cfu/g) (Table 4).

Table 4. Microbiological status of dried spices and herbs according to Recommendation 2004/24/EC¹³ and ESA¹² specifications from retail premises in the UK

Microorganism	Product Identification	Number of samples	Analysis results		
			Satisfactory	Acceptable	Unsatisfactory
<i>Salmonella</i> spp.	<i>Capsicum</i> spp.	534	528	0	6
	<i>Piper</i> spp.	327	323	0	4
	Other spice	1229	1218	0	11
	Herbs	743	733	0	10 [£]
<i>Bacillus cereus</i>	<i>Capsicum</i> spp.	534	512	15	7
	<i>Piper</i> spp.	327	299	25	3
	Other Spices	1229	1101	120	8 [¥]
	Herbs	743	685	56	2
<i>Clostridium perfringens</i>	<i>Capsicum</i> spp.	534	512	22	0
	<i>Piper</i> spp.	327	324	2	1
	Other spices	1229	1200	25	4 ^{£¥}
	Herbs	743	696	42	5
<i>Escherichia coli</i>	<i>Capsicum</i> spp.	534	520	12	2
	<i>Piper</i> spp.	327	316	6	5
	Other spices	1229	1188	28	13
	Herbs	743	664	40	39
Enterobacteriaceae			Analysis results		
			£100 cfu/g	>100 cfu/g	
	<i>Capsicum</i> spp.	534	295	239	
	<i>Piper</i> spp.	327	199	128	
	Other spices	1229	803	425	
	Herbs	743	275	469	

£ One sample detected with *Salmonella* spp. also had unsatisfactory levels of *C. perfringens*

¥ One sample had unsatisfactory levels of both *B. cereus* and *C. perfringens*

***Salmonella* spp. present in dried herbs and spices**

Salmonella spp. was detected in 1% (2/132) of production batches and 1% (31/2833) of samples collected from retail establishments. Details of the *Salmonella* serotypes recovered from the dried spices and herbs are provided in Table 5. Seventeen different *Salmonella* serotypes were obtained from 16 different types of dried herbs and spices. Twenty percent (6/30) of the referred isolates were identified as *S. Senftenberg*.

Table 5. *Salmonella* spp. isolates from dried spices and herbs

Type of Sample	Number of Batches/ Samples	<i>Salmonella</i> sero /phage types (resistance to antimicrobial drugs*)
Dried Spices:		
Cumin	3	S. Carcass (SuTTm) S. Montevideo <i>Salmonella</i> spp. [□]
Fennel	1	S. Edinburgh
Garam masala	1	S. Clifton
Chilli	3	S. Typhimurium RDNC [†] S. Senftenberg S. Aequartoria
Black pepper	5	S. Senftenberg S. Poona S. Infantis S. Schwarzengrund S. Saint- Paul
Turmeric	4	S. Senftenberg (2) S. Friedenau S. Tennessee
Cayenne	2	S. Hvitvingfoss <i>Salmonella</i> spp. [□]
All spice	1	S. Senftenberg
Cinnamon	1	S. Senftenberg
Curry	1	S. Montevideo
Okra	1	S. Hato
Dried Herbs:		
Coriander	6	S. Derby S. Montevideo (ACr) S. Infantis (SSuSpTFu) S. Agona (2), S. Typhimurium RDNC [†]
Fenugreek	1	S. Mbandaka
Oregano	1	<i>Salmonella</i> spp. [□]
Mint	1	S. Typhimurium DT [‡] 193
Sage	1	S. Agona

* Key to antimicrobial drugs: A, Ampillicin; C, Ciprofloxacin; Cr Cephradine, Fu, Fulphonamide S, Steptomycine; Su, Sulphonamides; T, Tetracycline; Tm, Trimethoprim

□ Isolates not referred for further typing

† Reacted with the phage set but did not confirm to a recognised phage type

‡ DT, definitive phage type

***Bacillus* spp. and *B. cereus* isolated at $\geq 10^5$ cfu/g from dried spices and herbs**

Of the 294 *Bacillus* spp. and *B. cereus* isolates at $\geq 10^5$ cfu/g (obtained from 227 samples; ranged from 1.0×10^5 – 6.9×10^9 cfu/g) referred for characterisation, most were (58%; 169/294) shown to be *B. subtilis*, and the remainder to be *B. pumilus*, *B. cereus* and *B. licheniformis*. (Table 6). Annex 1 provides further details about the types of *Bacillus* spp. isolated from specific spice or herb types.

Table 6. *B. cereus* and other *Bacillus* spp. isolated at $\geq 10^5$ cfu/g from dried spices and herbs

Dried spices	Number of Isolates*
<i>B. amyloliquefaciens</i>	1
<i>B. cereus</i>	37
<i>B. licheniformis</i>	32
<i>B. pumilus</i>	45
<i>B. subtilis</i>	166
Dried herbs	
<i>B. cereus</i>	8
<i>B. licheniformis</i>	1
<i>B. pumilus</i>	1
<i>B. subtilis</i>	3

* 67 samples had more than one type of *Bacillus* spp. present.

Enterobacteriaceae at ≤ 100 cfu/g present in dried herbs and spices

Forty five percent (59/132) of production batches and 56% (1572/2833) of retail spice and herb samples had Enterobacteriaceae levels below at ≤ 100 cfu/g. In this study the Enterobacteriaceae count was used as an indicator for possible irradiation or heat treatment of spices and herbs. Samples that had an Enterobacteriaceae count of ≤ 100 cfu/g were therefore considered as possibly having been irradiated or submitted to heat treatments.

Product Information in relation to microbiological quality

Product Detail

Using the classification of dried herbs and spices within Recommendation 2004/24/EC¹³ (Table 7), 18% of batches/samples were categorised as *Capsicum* spp., 12% were *Piper* spp., 44% were other spices, and 26% were dried herbs. The proportion of herb samples of unsatisfactory microbiological status was higher (6%) when compared to all other spice types (3-4% ($p=0.0013$)) (Table 7). The proportion of sage and basil samples of unsatisfactory status was significantly higher (19% and 13%, respectively) when compared to other dried herb types (0-8%) ($p<0.0001$) (Table 7).

Table 7. Microbiological status of dried spices and herbs according to Recommendation 2004/24/EC¹³ and ESA¹² specifications in relation to type of spices or herbs

Type of Spice or Herb	No Batches and Samples		No Batches and Samples of Unsatisfactory Quality (%)
	(n= 2965)	%	
<i>Capsicum</i> spp. (n= 552)	552	18%	17 (3%)
Chilli	260	47%	9 (3%)
Cayenne	78	14%	7 (9%)
Paprika	214	39%	1 (0.5 %)
<i>Piper</i> spp. (n=355)	355	12%	15 (4%)
Black pepper	243	8	13 (5%)
White pepper	70	2	2 (3%)
Green pepper	8	0.1	-
Other (mixed, red peppers, flower pepper)	34	1	-
Other Spices (n= 1292)	1292	44%	42 (3%)
Aniseed	47	2	1 (2%)
All spice	22	1	2 (9%)
Cinnamon	135	5	1 (1%)
Cumin	166	6	4 (2%)
Fennel	40	1	1 (3 %)
Garam Masala	92	3	6 (7%)
Ginger	130	4	5 (4%)
Mace	9	0.3	-
Mustard	38	1	-
Nutmeg	92	3	-
Saffron	2	0.1	-
Turmeric	163	6	10 (6%)
Other (tikka/ tandori masala, coriander powder)	356	11	12 (3%)
Dried Herbs (n= 766)	766	26%	48 (6%)
Basil	71	2	9 (13%)
Bay	26	1	-
Coriander	148	5	12 (8%)
Oregano	67	2	3(4%)
Parsley	72	2	1 (1%)
Rosemary	58	2	-
Sage	48	2	9 (19%)
Thyme	50	2	-
Tarragon	28	1	-
Other (dill, fenugreek, lemongrass, coltsfoot)	198	7	14 (7%)

Of the spice and herb samples collected, more than half were ground types (58%, 1721/2965) (Table 8). Significantly, a higher proportion of spices and herbs collected as flakes (9%) were of unsatisfactory microbiological quality compared with samples that were whole (2%), ground (2%) or in other formats (3%, i.e. granules, pieces) ($p < 0.0001$) (Table 8).

More than three-quarters (2277/2695; 77%) of the samples were pre-packed (Table 8). There was no difference in the proportion of spices and herbs collected as pre-packed packs (4%) or from opened packs at the premises (4%) that were of unsatisfactory microbiological status (Table 8).

The condition of most (2666/2965; 90%) spices and herbs sampled were recorded as ready-to-use (Table 8). There was no difference in the proportion of spices and herbs collected that were 'ready-to-use' (4%) or destined for further processing (4%) that were of unsatisfactory microbiological status (Table 8).

For most samples collected (2711/2965; 91%) there was no information to determine whether they had received a processing treatment or not (Table 8). However, 2% (74) of samples were labelled as having had received a processing treatment, while 6% (180) had not (Table 8). A higher proportion (11/180; 6%) of samples that had not received any treatment were of unsatisfactory microbiological status compared to those that had (2/74; 3%), although this finding was not statistically significant (Table 8) ($p=1.0000$).

Of the samples labelled as having had a treatment, 53% (39/74) were treated with hot air, 16% were steam treated, and additionally 31% were freeze-dried (Table 8). Significantly a greater proportion of samples that had been freeze-dried (9/23; 39%) were of unsatisfactory microbiological status compared to those that had received hot air (1/39; 3%) or steam (1/12; 8%) treatment (Table 8) ($p = 0.0003$). Twenty-two percent (16/74) of samples were treated in the UK, 74% were treated before export to the UK, and for 4% this information was not known (Table 8).

There was no evidence of mould growth in the majority (2819/2965; 95%) of samples. (Table 8). A higher proportion (2/7; 28%) of samples that were of unsatisfactory status were contaminated with mould when compared with samples that were not (118/2819; 4%). However, it should be noted that the proportion of samples with evidence of mould growth was very small and that no statistical conclusions should be drawn from these results.

Table 8. Microbiological status of dried spices and herbs according to Recommendation 2004/24/EC¹³ and ESA¹² specifications in relation to product detail

Product Detail	No Batches and Samples		No Batches Samples of Unsatisfactory Quality (%)
	(n= 2965)	%	
Type of spice or herb: texture is tactile not format			
Whole	420	14	9 (2%)
Ground	1721	58	68 (2%)
Flakes	326	11	29 (9%)
Other (e.g. granules, pieces)	142	5	4 (3%)
Not recorded	356	12	12 (3%)
Type of packaging			
Open pack	655	22	29(4%)
Unopened / sealed pack	2277	77	91 (4%)
Not recorded	33	1	2 (3%)
Sample status			
Ready-to-use	2666	90	106 (4%)
Destined for processing as part of food manufacturing process	118	3.7	5 (4%)
Destined for packing in the UK	9	0.3	-
Not known	172	6	11 (6%)
Product been treated			
Yes	74	2	2 (3%)
No	180	6	11(6%)
Not known	2711	91	109(4%)
Treatment (n= 74)			
Hot air treatment	39	53	1 (3%)
Steam treatment	12	16	1 (8%)
Freeze dried	23	31	9 (39%)
Treated in UK (n= 74)			
Yes	16	22	1 (6 %)
Before export to the UK	55	74	1 (2%)
Not Known	3	4	-
Evidence of mould growth			
Yes	7	0.2	2 (28%)
No	2819	95	118 (4%)
Not known	139	4.8	2 (1%)

Country of Origin

A fifth (21%) of the spices and herbs collected were produced outside the EU, and of these 57% (356/619) were produced in India (Table 9). The proportion of samples produced by non-EU countries of unsatisfactory microbiological status was significantly higher (6%; 37/619) than those produced within the EU (1%; 4/307) (P=0.0006).

Table 9. Microbiological status of dried spices and herbs according to Recommendation 2004/24/EC¹³ and ESA¹² specifications in relation to the country of origin

Country of origin	Number of Batches and Samples		No. Batches and Samples of unsatisfactory quality (%)
	n = 2965	%	
UK	148	5	-
Other EU	159	5	4 (3%)
France	7	4	-
Germany	10	6	-
Spain	102	64	1 (1%)
Other (Greece, Bulgaria, Ireland, Italy and others)	40	26	3 (7.5%)
Non- EU	619	21	37 (6%)
Australia	5	1	-
Brazil	5	1	1 (20%)
Canada	15	2	-
China	65	11	2 (3%)
Egypt	36	6	6 (17 %)
Guatemala	15	2	-
India	356	57	16 (5%)
Indonesia	22	4	2 (9%)
Other (Iran, Jamaica, Japan, Korea, Mexico and others)	100	16	10 (10 %)
Produce of more than one country	136	5	19 (14%)
Not known	1903	64	62 (3%)

Premises details in relation to microbiological quality

Type of premises

Food premises

Almost half of the spices and herbs were collected from supermarkets (29%) and convenience stores (20%). Samples were also collected from market stalls (3%), delicatessens (5%) and catering premises (16%) (Table 10). Significantly a higher proportion of samples of unsatisfactory microbiological quality were collected from convenience stores compared with those from supermarkets ($p=0.0001$).

Production premises

Most (78%) production spices and herbs were collected from manufacturers (47%) and packing centres (31%). No significant difference was observed in the proportion of samples of unsatisfactory microbiological status collected from the different production premises (Table 10).

Additionally 40 batches of dried herbs and spices were collected at border inspection posts (BIPs), with four (10%) of the batches found to be of unsatisfactory microbiological status.

Food Hygiene Inspections

Forty-four percent of samples were collected from premises categorised as inspection rating Category C (inspected every 18 months) (Table 10). The proportion of samples of unsatisfactory status was similar from premises with inspection rating categories A to F (ranging from 2-5%).

The majority of samples were obtained from premises categorised in consumer at risk score 5 (few numbers of customers, 59%) and 10 (intermediate numbers of customers, 18%) (Table 10). The proportion of samples of unsatisfactory status was similar from premises rating with different consumer at risk scores (ranging from 2-5%) (Table 10) ($P= 0.6755$)

Most samples (67%) were collected from premises that had a confidence in management score of 5 (moderate confidence in management/control systems 30%) or 10 (some confidence in management/control systems, 37%) (Table 10). Significantly a greater number of spices and herbs of unsatisfactory status were obtained from premises with high scores (13%, i.e. little or no confidence in the management) compared to those with a low score (10%, i.e. some to high confidence in management) (Table 10) ($p=0.0001$).

Table 10. Microbiological status of dried spices and herbs according to Recommendation 2004/24/EC¹³ and ESA¹² specifications in relation to premises details

Premises Details		No. Batches and Samples		No batches and samples of Unsatisfactory Quality (%)
		(n= 2922)	%	
Retail Premises Type (n=2833)		2833	97	111 (4%)
Supermarket		826	29	15 (2%)
Convenience store		598	21	34 (6%)
Market stall		88	3	4(5%)
Delicatessen		144	5	7 (5%)
Catering premises		446	16	15(3%)
Other (Butcher shops, farm shops, health food shops, specialist shops)		730	26	36 (5%)
Production Premises Type (n=89)		89	3	6 (7%)
Importers		14	16	2 (14%)
Packing centre		28	31	2 (7%)
Wholesalers		5	6	1 (20%)
Manufacturers		42	47	1 (2%)
Inspection Rating Category				
Category	Minimum Frequency of inspection			
A	At least every 6 months	61	2	1 (2%)
B	At least every 12 months	287	10	12 (4%)
C	At least every 18 months	1316	44	53 (4%)
D	At least every 2 years	451	15	15 (3%)
E	At least every 3 years	286	10	10 (3%)
F	At least every 5 years	78	3	4 (5%)
Not recorded		443	16	22(7%)
Consumer at Risk Score				
0 (Very few)		46	2	2 (4%)
5 (Few)		1721	59	72 (4%)
10 (Intermediate)		522	18	12 (2%)
15 (Substantial)		119	4	6 (5%)
Not Recorded		514	18	25 (5%)
Confidence in Management				
0 (High)		161	5	5 (3%)
5 (Moderate)		875	30	24 (3%)
10 (Some)		1096	37	49 (4%)
20 (Little)		243	8	12 (5%)
30 (None)		25	1	2 (8%)
Not Recorded		522	18	25 (5%)
Hazard Analysis Systems				
In place and documented		1100	38	35 (3%)
In place and not documented		590	20	34 (6%)
Not in place		651	22	25 (4%)
Not Recorded		581	20	19 (3%)
Management Food Hygiene Training				
Received training and attended		1853	63	65 (3%)
❖ Basic 6 hour course		1290	69	47 (4%)
❖ Intermediate course		261	14	7 (3%)
❖ Advanced course		93	5	3 (3%)
❖ Other recognised course		209	11	2 (1%)
No training		565	19	29 (5%)
Not Recorded		504	19	23 (5%)

Hazard analysis systems

Fifty eight percent of samples were collected from premises that had hazard analysis in place (Table 10). A similar proportion of spices and herbs of unsatisfactory status were collected from premises that had hazard analysis in place (3%) compared to premises that did not (4%).

Food Hygiene Training

Sixty three percent of samples were collected from premises whose managers had received some form of training (Table 10). A higher proportion of samples of unsatisfactory quality were from premises where the manager had not received food hygiene training (5%) compared to those that had (3%), although this finding was not significant ($p=0.0185$).

Discussion

Dried spices and herbs are used in a variety of ways by manufacturers, caterers and the public, and microbiological concerns of these products are governed by end use. Although spices and herbs are not major contributors to foodborne disease, a potential hazard exists, particularly if the spice or herb is added at the end of cooking or added to prepared foods without further cooking^{7,21}. The establishment of microbiological specifications, such as those by Codex¹⁰ and ESA¹², are related to the final use of spices and herbs and require that *Salmonella* should be absent in treated spices and herbs and free from pathogenic microorganisms in levels that may represent a hazard to health. This study has shown that the vast majority of dried spices and herbs sampled at retail (96%) and (92%) production premises in the UK were of satisfactory or acceptable microbiological quality according to microbiological criteria in the EC Recommendation 2004/24/EC¹³ and ESA specifications¹².

Salmonella spp. have been found in a wide variety of herbs and spices with rates of contamination ranging between 0.6% and 14%^{3,4,6,21-23}. The prevalence of *Salmonella* spp. in dried spices and herbs in the UK in 2004 (1%) was similar to that found in Ireland (0.9%; in 2004)²², Japan (0.7%; in

2006)⁴, and Austria (0.6%; in 1994)³, but lower than that previously found in Australia (2.0%; in 1986)⁶, the USA (6.5%; in 1987)²³, and the UK (12.3%; in 1996, C Little, HPA pers comm). The presence of *Salmonella* spp. is of particular concern when herbs and spices are added to ready-to-eat foods, and contaminated spices have been responsible for outbreaks of salmonellosis⁵⁻⁸.

Spore forming bacteria (*B. cereus*, *C. perfringens*) that are capable of causing foodborne disease when ingested in large numbers are frequently found in spices and herbs, but usually at low levels²¹. Levels of *B. cereus* at $\geq 10^4$ cfu/g in herbs and spices in the UK in 2004 (1%) were lower than that previously found in the UK (1.9%; in 2003)⁷. *B. cereus* at $\geq 10^4$ cfu/g has also been reported in a variety of herbs and spices sampled in Australia, Hungary, Ireland, Italy, The Netherlands, and the USA, with the proportion ranging between 0% to 6.5%^{6,22,24-28}. *B. subtilis* is commonly found in many spices²¹ and of the *Bacillus* spp. at high levels ($\geq 10^5$ cfu/g) in this study, over half (58%) were identified as *B. subtilis*. However, *B. subtilis* has only occasionally been a cause of food poisoning linked to spices^{7,29}. *C. perfringens* has also been found in several spices usually at < 500 cfu/g and rarely $\geq 10^3$ cfu/g²¹. *C. perfringens* at $\geq 10^3$ cfu/g in herbs and spices in the UK in 2004 was 0.4%. In other studies *C. perfringens* at $\geq 10^3$ cfu/g ranged from 0% to 7.6%^{6,21,27,30}. Because spores of the above organisms may survive cooking temperatures and may multiply to high levels in foods held at room temperatures, a risk of food poisoning by these pathogens may be created when no proper time-temperature conditions for heating, cooling and storage of foods seasoned with spices or herbs are applied.

All spices and herbs are susceptible to microbial contamination. The prevention of microbial contamination in dried herbs and spices lies in the application of good hygiene practices during growing, harvesting and processing from farm to fork, and effective decontamination. Codex has produced a Code of Hygienic Practice for dried herb and spice suppliers¹⁰ to help minimize microbial food safety hazards. The EC Regulation on the hygiene of foodstuffs³¹ advocates the principles of HACCP and the

establishment of in-process controls to ensure product integrity, rather than reliance on end-product testing for compliance with specifications. In the spice and herb industry the strengthening of HACCP systems that encompass all stages of production, processing and distribution will serve to further enhance the microbial safety of these products. In addition, educating food handlers and consumers as to the importance of correct food handling practices will also help to prevent foodborne illness due to contaminated spices and herbs.

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Annex 1

Table 1. *B. cereus* and *Bacillus* spp. isolated from dried spices and herbs
 (*Total No. = 626; of these 399 samples were not sent for further typing)

Type of Sample	No. of Batches/ Samples	<i>Bacillus</i> spp.*
Dried Spices:		
All spice	8	<i>Bacillus</i> spp (5)* <i>B. subtilis</i> <i>B. licheniformis</i> <i>B. licheniformis</i> / <i>B. subtilis</i>
Aniseed	1	<i>B.subtilis</i>
Chill (Powdered, flakes and Ground)	120	<i>Bacillus</i> spp. (68)* <i>B. licheniformis</i> (3) <i>B. pumilus</i> (3) <i>B. cereus</i> (10) <i>B. subtilis</i> (30) <i>B. amyloliquefaciens</i> , <i>B. subtilis</i> / <i>B. licheniformis</i> <i>B. subtilis</i> / <i>B. pumilus</i> (3) <i>B. subtilis</i> / <i>B. pumilus</i> / <i>B. licheniformis</i>
Coriander (Ground)	9	<i>Bacillus</i> spp. (6)* <i>B. cereus</i> <i>B. pumilus</i> / <i>B. licheniformis</i> <i>B. pumilus</i> / <i>B. cereus</i>
Cumin	9	<i>Bacillus</i> spp. (6)* <i>B. licheniformis</i> <i>B. subtilis</i> <i>B. subtilis</i> / <i>B. pumilus</i>
Curry powder	45	<i>Bacillus</i> spp.(28)* <i>B. pumilus</i> <i>B. cereus</i> <i>B. subtilis</i> (7) 3 x <i>B.subtilis</i> / <i>B. pumilus</i> (3) <i>B. subtilis</i> / <i>B. licheniformis</i> <i>B. subtilis</i> / <i>B. licheniformis</i> / <i>B. cereus</i> (2) <i>B. subtilis</i> / <i>B. licheniformis</i> / <i>B. pumilus</i> / <i>B. cereus</i> <i>B. cereus</i> / <i>B. subtilis</i>
Garam masala	53	<i>Bacillus</i> spp. (38)* <i>B.subtilis</i> (7) <i>B. cereus</i> <i>B. pumilus</i> (2) <i>B. licheniformis</i> , <i>B. cereus</i> / <i>B. subtilis</i> <i>B. subtilis</i> / <i>B. pumilus</i> <i>B. pumilus</i> / <i>B. Licheniformis</i>
Ginger	18	<i>Bacillus</i> spp. (9)* <i>B.cereus</i> <i>B. subtilis</i> (4)

		<i>B. pumilus</i> <i>B. subtilis</i> / <i>B. pumilus</i> (3)
Mixed spices	7	<i>Bacillus</i> spp. (6)* <i>B. pumilus</i> / <i>B. cereus</i>
Nutmeg	2	<i>Bacillus</i> spp. (2)*
Paprika	49	<i>Bacillus</i> spp. (27)* <i>B. subtilis</i> (13) <i>B. cereus</i> <i>B. pumilus</i> <i>B. pumilus</i> / <i>B. cereus</i> / <i>B. subtilis</i> <i>B. subtilis</i> / <i>B. pumilus</i> / <i>B. licheniformis</i> <i>B. subtilis</i> / <i>B. licheniformis</i> <i>B. subtilis</i> / <i>B. pumilus</i> (4)
Pepper (black)	134	<i>Bacillus</i> spp. (96)* <i>B. cereus</i> (2) <i>B. subtilis</i> (21) <i>B. pumilus</i> / <i>B. subtilis</i> (8) <i>B. pumilus</i> / <i>B. circulans</i> <i>B. pumilus</i> / <i>B. licheniformis</i> <i>B. subtilis</i> / <i>B. cereus</i> (2) <i>B. subtilis</i> / <i>B. pumilus</i> / <i>B. cereus</i> <i>B. subtilis</i> / <i>B. pumilus</i> / <i>B. licheniformis</i>
Pepper (red)	3	<i>Bacillus</i> spp. (2)* <i>B. subtilis</i> / <i>B. pumilus</i>
Pepper (White)	1	<i>Bacillus</i> spp. *
Pepper (mixed)	3	<i>Bacillus subtilis</i> (3)
Saffron	1	<i>Bacillus</i> spp.*
Trumeric	68	<i>Bacillus</i> spp. (28)* <i>B. subtilis</i> (23) <i>B. cereus</i> <i>B. pumilus</i> (2) <i>B. licheniformis</i> (4) <i>B. subtilis</i> / <i>B. licheniformis</i> (5) <i>B. licheniformis</i> / <i>B. cereus</i> <i>B. cereus</i> / <i>B. subtilis</i> <i>B. subtilis</i> / <i>B. pumilus</i> <i>B. subtilis</i> / <i>B. licheniformis</i> / <i>B. pumilus</i>
Other	80	<i>Bacillus</i> spp. (48)* <i>B. subtilis</i> (14) <i>B. pumilus</i> (3) <i>B. cereus</i> (8) <i>B. pumilus</i> / <i>B. cereus</i> (2) <i>B. pumilus</i> / <i>B. subtilis</i> (2) <i>B. pumilus</i> / <i>B. licheniformis</i> <i>B. licheniformis</i> / <i>B. subtilis</i> <i>B. subtilis</i> / <i>B. licheniformis</i> / <i>B. cereus</i>
Dried Herbs:		
Basil	5	<i>Bacillus</i> spp. (3)* <i>B. subtilis</i> <i>B. pumilus</i>

Dill	1	<i>Bacillus</i> spp.*
Fenugreek	1	<i>Bacillus</i> spp.*
Mint	2	<i>Bacillus</i> spp. (2)
Sage	1	<i>Bacillus</i> spp.*
Tarragon	1	<i>Bacillus</i> spp.*
Thyme	1	<i>B. lichenformis</i>
Other (Lemon grass)	2	<i>B. subtilis</i> <i>B. subtilis/B .cereus</i>

Annex 2: Participating Laboratories and Local Authority Food Liaison Groups

Table I: Participating HPA and HPA Collaborating Laboratories and number of samples

HPA Region	Laboratory Name	No. of Samples
East	Chelmsford	163
	Norwich	104
London	London FWEM ¹	186
South East	Ashford	94
	Brighton	193
	WEMS ²	273
West Midlands	Birmingham	75
	Coventry	169
	Shrewsbury & Telford	56
	Stoke	59
	Hereford	20
North West	Chester	52
	Preston	240
	Carlisle	26
North East, Yorkshire & the Humber	Hull	95
	Leeds	82
	Newcastle	53
	Sheffield	262
South West	Bristol	92
	Exeter	84
	Plymouth	25
	Truro	62
East Midlands	Leicester	94
	Lincoln	210
Total		2769

1, London Food, Water & Environmental Microbiology Laboratory

2, Wessex Environmental Microbiological Services

Table II: Participating Other Laboratories and number of samples

Nation	Laboratory	No. of Samples
England	Kings Lynn & West Norfolk	12
	Worcester Scientific Services	16
Northern Ireland	Belfast City Hospital	60
	Royal Alexandria, Paisley	4
Scotland	Aberdeen City Council Public Analysts	32
	Dundee Scientific Services	4
	Glasgow Scientific Services	9
Wales	Bangor	35
	Rhyl	24
Total		196

Table III: Participating Food Safety Liaison Groups and number of samples

Local Authority Food Liaison Group	No. of Samples
Berkshire	44
Buckinghamshire	23
Cambridgeshire	67
Cheshire	23
Cornwall	44
Cumbria	43
Derbyshire	167
Devon	76
Dorset	25
Durham	6
East Sussex	67
Essex	78
LFCG ¹ Greater London NE Sector	53
LFCG Greater London NW Sector	59
LFCG Greater London SE Sector	31
LFCG Greater London SW Sector	33
Greater Manchester	106
Hampshire & Isle Of Wight	167
Hereford & Worcester	36
Hertfordshire & Bedfordshire	10
Humberside	83
Kent	94
Lancashire	112
Leicestershire	89
Lincolnshire	94
Merseyside	24
North Yorkshire	40
Northamptonshire	87
Northern Ireland Food Group ²	60
Norfolk	97
Nottinghamshire	108
Oxfordshire	18
Scottish Food Enforcement Liaison Committee ³	55
Shropshire	33
Somerset	34
South West Yorkshire	103
Staffordshire	59
Suffolk	48
Surrey	77
Tyne & wear	34
Wales North Group	58
Wales South West Group	6
Warwickshire	84
West Midlands	102
West of England	58
West Sussex	54
West Yorkshire	54
Wiltshire	42
Total	2965

1, London Food Co-ordinating Group

2, Northern Ireland Food Group comprises of the Eastern, Northern, Southern and Western Groups

3, SFELG comprises of Central Scotland, Fife & Tayside, Lothian & Scottish Borders, North Scotland, and West of Scotland